

# Docetaxel and a novel bcl-2 antisense oligonucleotide for enhanced apoptosis in prostate cancer cells: A potential therapeutic strategy

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## ABSTRACT

**Background:** Prostate cancer is a leading cause of morbidity and mortality among men. Overexpression of Bcl-2 proteins is linked to prostate cancer progression, suggesting that targeting Bcl-2 could enhance therapeutic efficacy. Therefore, Bcl-2 inhibition may potentiate anti-tumor activity in prostate cancer. **Methods:** We designed a novel antisense oligonucleotide (ASO) derived from G3139 and evaluated its anti-proliferative effects in LNCaP and PC3 prostate cancer cell lines. The apoptotic effects of docetaxel (DTX) and the ASO, administered alone or in combination, were assessed by real-time PCR and Annexin V-FITC/PI flow cytometry. We hypothesized that the combination therapy would be more effective than either agent alone. **Results:** The ASO significantly down-regulated Bcl-2 mRNA and inhibited proliferation in both cell lines, as shown by real-time PCR. Flow cytometry revealed an increase in early apoptotic cells in all treated groups. Co-treatment with the ASO and DTX produced the highest apoptotic rates (75–85 %). **Conclusions:** The novel ASO effectively reduces Bcl-2 expression and sensitizes prostate cancer cells to docetaxel, permitting lower DTX doses. The combination therapy induced 75–85 % apoptosis in LNCaP and PC3 cells, suggesting an improved strategy that may limit high-dose DTX-associated resistance and toxicity. **Key words:** Bcl-2, G3139, Antisense oligonucleotide (ASO), Docetaxel (DTX), Prostate cancer

## INTRODUCTION

Prostate cancer is the second most common cancer and the fifth leading cause of cancer-related death in men. The incidence and mortality rates of prostate cancer are strongly related to age, with the highest rates observed in men over 65 years. Other risk factors include ethnicity, genetics, and family history<sup>1–4</sup>. The progression and development of prostate cancer is largely related to the number of genetic abnormalities that affect not only the androgen receptor but also the regulation of apoptotic pathways<sup>5</sup>. Prostate cancer growth is initially androgen-dependent, but later becomes androgen-independent and is accompanied by increased expression of anti-apoptotic genes, leading to metastasis and higher mortality. Studies show that altered expression of apoptosis-regulating proteins contributes to therapy-resistant prostate cancer. In addition, defects in apoptotic pathways promote cancer-cell survival and resistance to chemotherapy drugs<sup>6–9</sup>.

The BCL-2 family plays an essential role in the regulation of cellular apoptosis. BCL-2 is a key anti-

apoptotic gene in this pathway<sup>10</sup>. The BCL-2 protein is anti-apoptotic, and its expression is directly related to the progression of many cancers. Overexpression of this gene is associated with resistance to several therapeutic stimuli, including androgen deprivation, chemotherapy, and radiotherapy<sup>11–14</sup>. Overexpression of the BCL-2 protein, common in malignant cells, promotes survival and drug resistance but also presents opportunities for targeted therapies that selectively eliminate these cells<sup>15</sup>. Thus, combining BCL-2 inhibition with other treatments may enhance cancer-cell killing and limit drug resistance and metastasis.

A common way to treat prostate cancer is chemotherapy; docetaxel (DTX) is often referred to as the first-line treatment. It stabilizes microtubules, thereby inducing cancer-cell death<sup>16,17</sup>. Another approach involves antisense oligonucleotides (ASOs), used alone or in combination with other agents. ASOs are chemically modified single-stranded oligodeoxynucleotides of 18–21 nt. These ASOs hybridize to their specific mRNAs, and the RNase H complex subsequently degrades the duplex.

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In other words, such ASOs can prevent translation of the target gene by binding to the mRNA<sup>18-20</sup>. Oblimersen (G-3139; Genasense; Genta Inc.) is an ASO that inhibits translation of BCL-2<sup>21</sup>.

The use of antisense oligonucleotides targeting BCL-2 in combination with docetaxel reduced the IC<sub>50</sub> of DTX compared with DTX alone. This reduction helps minimize toxicity to normal cells<sup>22</sup>. Furthermore, the combination therapy sensitized cancer cells to DTX and decreased their drug resistance<sup>22,23</sup>. As high doses of chemotherapy agents typically induce resistance in cancer cells, diminishing the dose can be a strategy to overcome drug resistance effectively<sup>22</sup>.

We designed a novel antisense oligonucleotide, based on the Ensembl BCL-2 mRNA sequence and the G3139 antisense oligonucleotide, to investigate its apoptotic effects on LNCaP and PC3 prostate-cancer cell lines, alone and in combination with docetaxel. We assessed apoptosis by real-time RT-PCR and flow cytometry, using Lipofectamine for delivery of docetaxel and the novel ASO. We hypothesized that targeting BCL-2 with ASO would enhance docetaxel-induced apoptosis by reducing the IC<sub>50</sub> of DTX.

Studies have shown that combining low-dose docetaxel with chemosensitizers can induce effective cytotoxicity in prostate-cancer cells while sparing normal prostate epithelial cells<sup>24</sup>. Clinically, reduced-dose docetaxel regimens have also demonstrated comparable efficacy with significantly decreased systemic toxicity<sup>25</sup>. In a rat study, histological examination of vital organs, including the prostate, two weeks after ASO treatment revealed no toxicity<sup>26</sup>.

**METHODS**

In this study, we designed a novel 22-mer antisense oligonucleotide (ASO) based on the Ensembl BCL-2 mRNA sequence and the previously reported 18-mer ASO G3139. We then evaluated and compared the pro-apoptotic effects of docetaxel (DTX) and the new ASO in LNCaP and PC3 prostate cancer cell lines using real-time PCR and flow cytometry.

**Cell Culture**

Prostate cancer cell lines LNCaP and PC3 were purchased from the Pasteur Institute (Tehran, Iran). Cells were cultured in RPMI-1640 supplemented with 10 % FBS and maintained at 37 °C in 5 % CO<sub>2</sub>. Cells were passaged at ~95 % confluence; medium was renewed every doubling time. The doubling times for LNCaP and PC3 were 72 h and 48 h, respectively.

**Docetaxel (DTX) Preparation**

Docetaxel (DTX) was obtained from Actover, dissolved in DMSO to 4 mg mL<sup>-1</sup> (10 mg in 2.5 mL) according to the manufacturer's instructions, aliquoted, and stored at -20 °C.

**Antisense Oligonucleotide Design and Preparation**

G3139 (oblimersen sodium, Genasense) is an 18-mer antisense oligonucleotides (5'-TCT CCC AGC GTG CGC CAT-3') complementary to codons 1-6 of the Bcl-2 mRNA<sup>27</sup>. Novel antisense oligonucleotides (ASO) designed according to the published nucleotide sequence of the bcl-2 mRNA in the Ensembl database ([https://asia.ensembl.org/Homo\\_sapiens](https://asia.ensembl.org/Homo_sapiens)) and based on the antisense oligonucleotide G3139.

The secondary structure of the bcl-2 mRNA predicted by minimum free energy (MFE) approach (<http://rna.tbi.univie.ac.at/cgibin/RNAWebSuite/RNAfold.cgi>). The sequence of ASO was 5'-GTTCTCCCAGCGTGCGCCATCC-3'. To enhance the stability of the antisense oligonucleotide at the target site within cells, we designed it with phosphorothioate modifications at both ends. It has 22 nucleotides, all 22 nucleotides have phosphorothioate modification but the first six nucleotides and the last six nucleotides also have 2'-O-(2-methoxyethyl) modification. It Purchased from Gorgon Gene Inhibition Biotechnology Company (Iran) (Table 1). The secondary structure of the bcl-2 mRNA address on minimum free energy (MFE) access with (<http://rna.tbi.univie.ac.at/cgibin/RNAWebSuite/RNAfold.cgi>) can exhibit in Figure 1.

**Binding Analysis**

The binding energies of the novel ASO and G3139 were compared *in silico*. A more negative  $\Delta G$  indicates higher duplex stability. G3139 displayed a  $\Delta G$  of -27.53 kcal mol<sup>-1</sup> (Figure 2 B), whereas the novel ASO reached -36.77 kcal mol<sup>-1</sup> (Figure 2 A), suggesting ~9 kcal mol<sup>-1</sup> stronger binding to BCL-2 mRNA.

**Primer Design for Real-Time PCR**

Real-time PCR primers were designed with Primer3 and Oligo 7 from the RefSeq sequences of GAPDH and BCL-2 (<https://ncbi.nlm.nih.gov/RefSeq>). GAPDH: forward 5'-CCT GCC GTC TAG AAA AAC CTG CCA-3', reverse 5'-CAG CGT CAA AGG TGG AGG AGT GGG-3' (156 bp). BCL-2: forward 5'-GAC GAC TTC TCC CGC CGC TAC-3', reverse

Table 1: Bcl-2 antisense oligonucleotide specifications

Sequence: 5'- *G2 -*T2-*T2-*C2-*T2-*C2-*C-*C-*A-*G-*C- *G-*T-*G-*C-*G-*C2-*C2-*A2-*T2-*C2-*C2 -3'	
Concentration: 100 μM	Synthesis scale: 0.20 μmol
GC%:68.2	MW: 6977 g/mol
Delivered: 349 μg-50.1 nmol	Synthesised: 10.5 OD
Quality Control: MALDI-TOF	Purification: HPLC

\*: phosphorothioate and 2: 2'-O-(2-methoxyethyl) is the modification of nucleotides

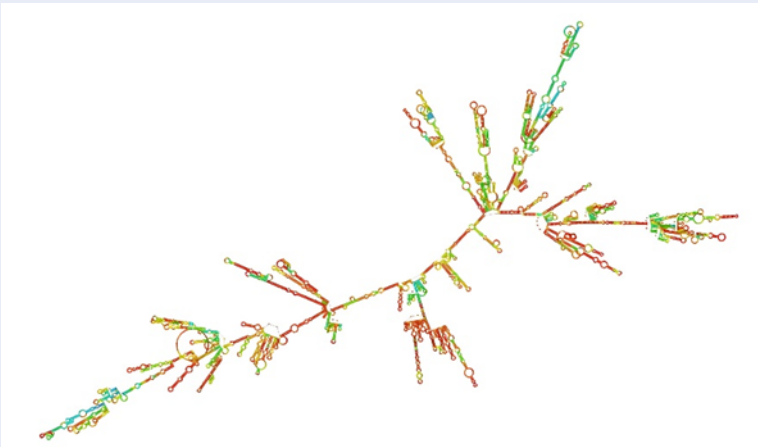


Figure 1: Prediction of secondary structure of mRNA bcl-2 based on MFE.

Table 2: Transfection and treatment of two cell lines with Antisense Oligonucleotide design and DTX

Cell lines	Control (48 and 72h)	Docetaxel (DTX) (48 and 72h)	New designed ASO (48 and 72h)	Docetaxel (DTX) and new ASO encapsulated with lipofectamine (48 and 72h)
LNCaP	*	*	*	*
PC3	*	*	*	*

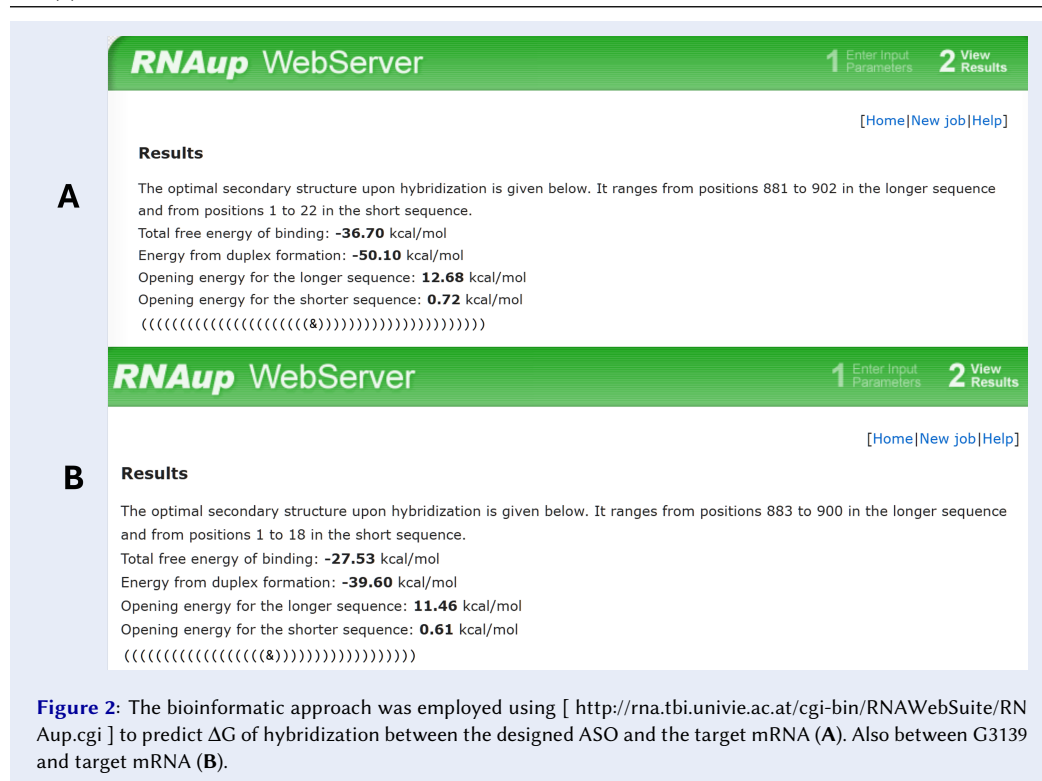
5'-TCC CCC AGT TCA CCC CGT CC-3' (133 bp). Despite the absence of a fluorescent label, significant BCL-2 down-regulation confirmed efficient delivery. Primer-BLAST confirmed specificity (Figure 3).

Transfection of Cells

LNCaP and PC3 cells at passage 6 were transfected with the ASO using Lipofectamine 2000 (Invitrogen) at a 1:1 (v/v) ratio (50 μL each) in serum- and antibiotic-free medium (Table 2). Final ASO concentrations were 125, 250, 500, and 1000 nM to determine the IC<sub>50</sub> for BCL-2 knock-down. After 30 min complex formation, the mixture was added to the cells and incubated for 48 h or 72 h in triplicate. The final Lipofectamine concentration was 25 μL mL<sup>-1</sup>.

RNA Isolation and cDNA Synthesis

Total RNA was isolated 48 h and 72 h post-transfection using TRIzol (Invitrogen) followed by DNase I treatment (Yekta Tajhiz, Iran). Quantity and purity were assessed spectrophotometrically (A<sub>20/20</sub>) and by agarose-gel electrophoresis, then verified with the BIOFACT RP101-050 kit. First-strand cDNA was synthesized with the YTA cDNA synthesis kit (Yekta Tajhiz).



### Analysis of BCL-2 mRNA Expression

Real-time PCR reactions (12  $\mu$ L) contained 1–2  $\mu$ L cDNA, 0.5  $\mu$ L each primer (5 pmol), 5  $\mu$ L RealQ Plus SYBR Green master mix (Amplicon, Denmark), and 4–5  $\mu$ L nuclease-free water. Cycling conditions were 95 °C/10 min; 45 cycles of 95 °C/30 s, 60 °C/30 s, 72 °C/30 s on an ABI StepOne system. Amplification efficiency ( $E = 10^{(-1/slope)}$ ) was calculated from standard curves (Figure 4). Relative expression was determined by  $\Delta\Delta C_t$  using GAPDH as the reference.

### Evaluation of Cell Cytotoxicity by MTT Assay

LNCaP and PC3 cells ( $5 \times 10^3$  cells/200  $\mu$ L) were seeded in 96-well plates. DTX cytotoxicity was measured by MTT assay after 48 h and 72 h. Concentration ranges were 2–400 nM (LNCaP) and 10–800 nM (PC3).  $IC_{50}$  values guided combination studies in which cells were first transfected with 125 or 250 nM ASO for 5 h, then treated with DTX at or below the  $IC_{50}$ . Absorbance at 570 nm was recorded. All experiments were performed in triplicate.

### Determination of Apoptosis by Flow Cytometry

Apoptosis was quantified with the Annexin V-FITC/PI Kit (MabTag, Germany) according to the

manufacturer's protocol. Cells were harvested after treatments defined by the MTT and qRT-PCR results, stained, and analyzed on a BD FACSArray with FlowJo software. Each condition was tested in triplicate.

### Statistical Analysis

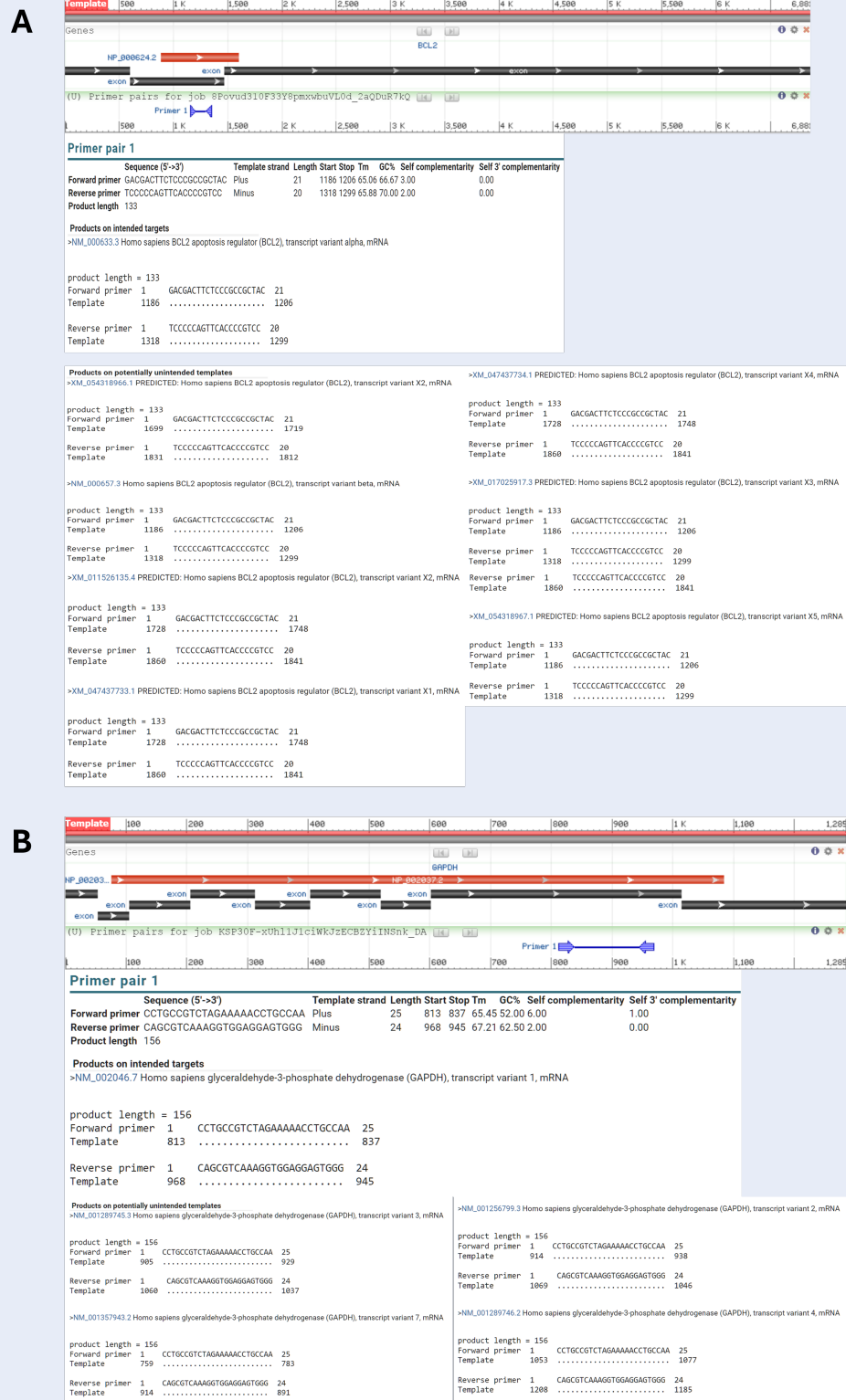
Data are presented as mean  $\pm$  SEM ( $n = 3$ ). One-way ANOVA followed by Student's t-test was used to assess significance. Graphs were generated with GraphPad Prism 8. PCR data were analyzed with LinRegPCR 11.0 and REST. Drug-interaction synergy was calculated by the Bliss independence model (Table 3).

## RESULTS

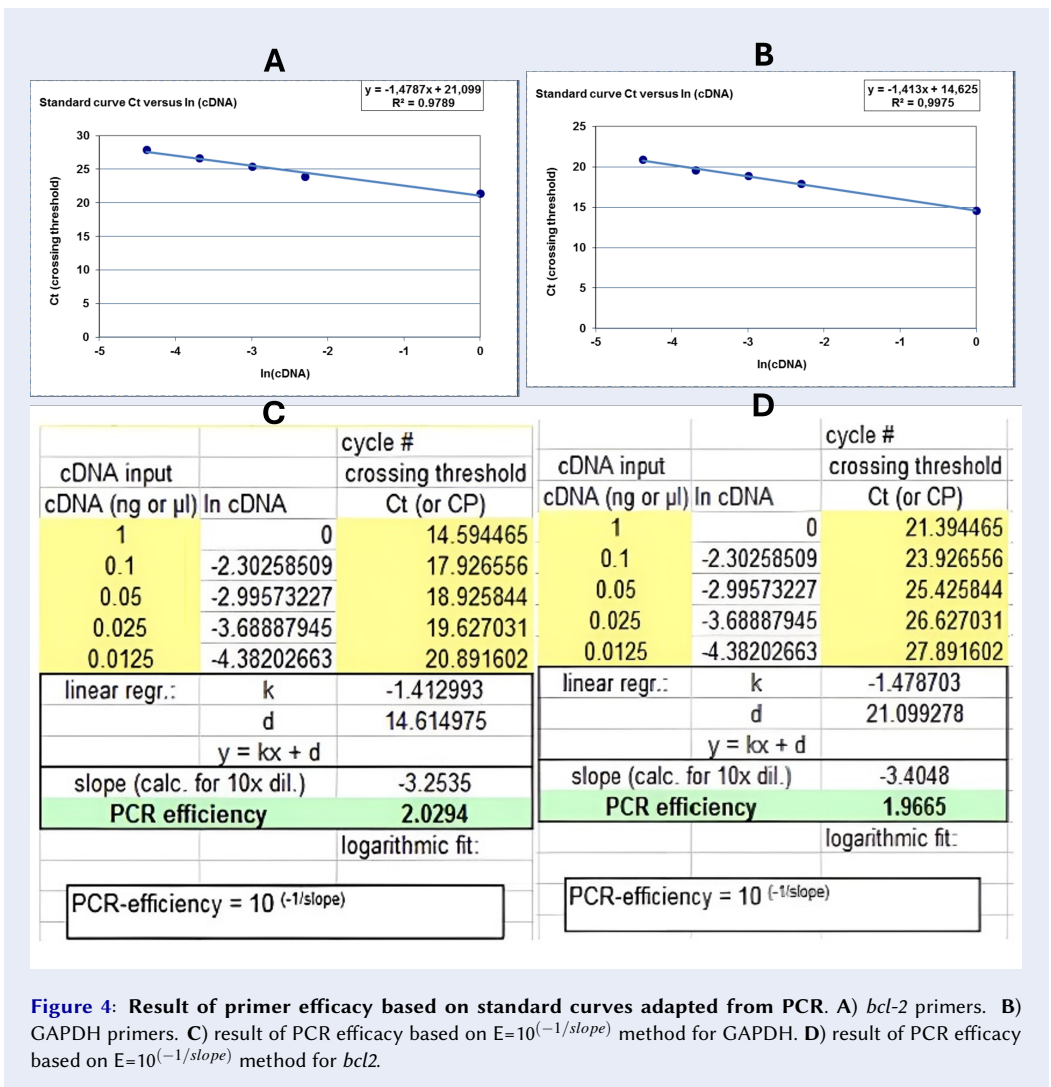
### Real-Time RT-PCR

To evaluate the effect of the newly designed ASO on BCL-2 expression in two prostate-cancer cell lines, we performed real-time RT-PCR as described in the Transfection section. BCL-2 mRNA levels were normalized to the housekeeping gene GAPDH. Down-regulation of BCL-2 was observed at both 48 h and 72 h with ASO concentrations of 250 nM and 125 nM, respectively (Figure 5).





**Figure 3:** Result of primer blast for A) *bcl-2* primers, and B) *GAPDH* primers in NCBI. *bcl-2* primers can produce 7 variants of *bcl-2* mRNA and *GAPDH* primers can produce 4 variants which means they designed for conserved regions of *bcl-2* mRNA and *GAPDH* mRNA.



MTT Assay

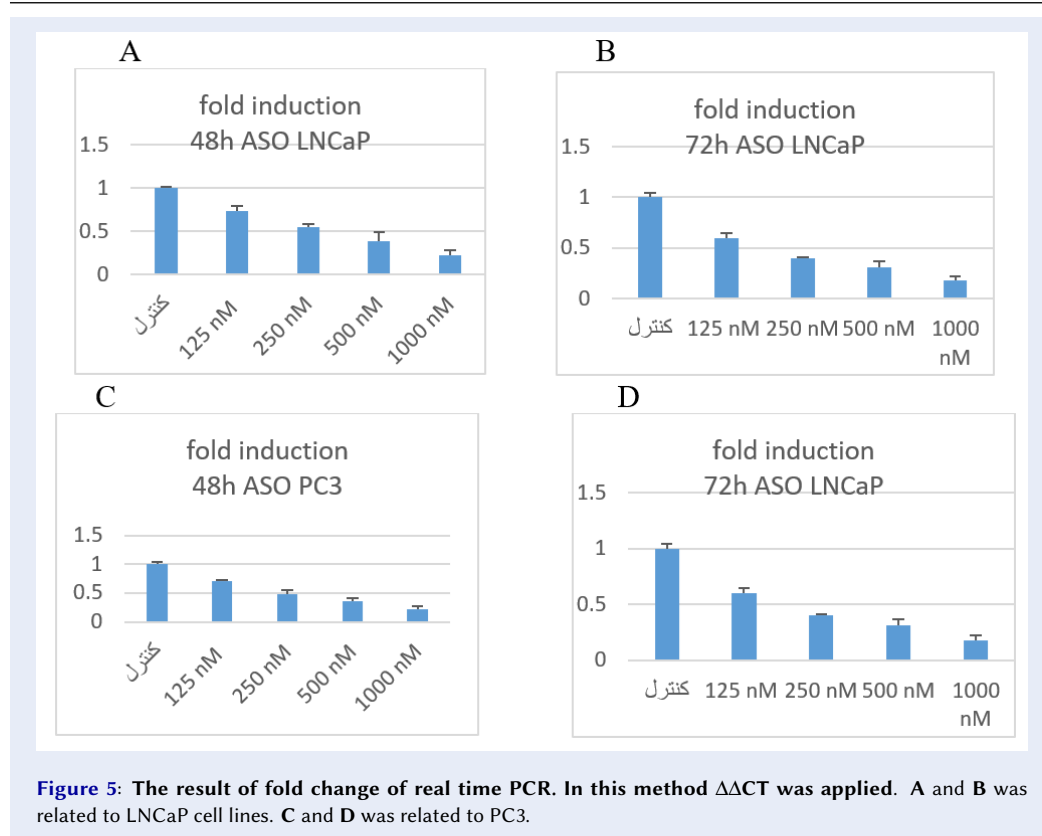
Cell viability decreased in a time- and dose-dependent manner. In the DTX-only group, after 48 h and 72 h, the IC<sub>50</sub> values were 135 nM and 63 nM for LNCaP cells and 89 nM and 57 nM for PC3 cells (Figure 6A–D). In the ASO-only group, the IC<sub>50</sub> values for LNCaP cells were 273 nM (48 h) and 189 nM (72 h), whereas for PC3 cells they were 309 nM (48 h) and 188 nM (72 h) (Figure 6E,F). In the ASO + DTX group, the IC<sub>50</sub> values for LNCaP cells were 75 nM (48 h) and 32 nM (72 h) (Figure 6G,H); for PC3 cells they were 48 nM (48 h) and 32 nM (72 h) (Figure 6 I,J).

In PC3 cell line in 48 h in the control group, *bcl-2* relative to the GAPDH was normal (100%). Nevertheless, in cells that had taken 125 nM, the expression of the *bcl-2* in relation to the GAPDH was almost

70%. In cells which had 500 nM ASO, the expression of *bcl-2* mRNA in ratio of GAPDH was approx 49%. The cells that had taken 500 nM, the expression of the *bcl-2* in relation to the GAPDH was almost 36%. In cells that had taken 1000 nM ASO, the expression of *bcl-2* was 21%.

Apoptosis

In LNCaP cells after 48 h, cell viability was 93 % in the control, 76 % with DTX alone, 74 % with ASO alone, and 16 % with the combination (Figure 7 A–D). In PC3 cells, viability was 91 % in the control, 49 % with DTX, 34 % with ASO, and 13 % with the combination (Figure 7 E–H). The combination exhibited a synergistic effect: the observed fractional inhibition (75 %) exceeded the Bliss-independence value (39 %) (Table 3). In PC3 cells, the combination lowered the



DTX  $IC_{50}$  and achieved 85 % apoptosis, confirming synergism.

## DISCUSSION

Prostate cancer is the second most common cancer in men. There are many risk factors for prostate cancer, including age, ethnicity, genetics and family history. Studies show that genetic abnormalities affect not only the androgen receptor but also the regulation of apoptosis<sup>1-5</sup>. Prostate cancer can increase the expression of anti-apoptotic genes; it can be immortal as long as the patient lives<sup>28</sup>. Peer-reviewed studies have reported that defects in apoptotic pathways can cause cancer-cell survival and resistance to chemotherapeutic agents<sup>29</sup>. BCL-2 plays an essential role in apoptosis<sup>30</sup>. Overexpression of this gene is associated with resistance to several anticancer therapies<sup>31</sup>.

It seems that apoptosis induction by chemotherapeutic agents is more effective when combined with BCL-2 silencing. BCL-2 silencing can occur at the mRNA level. Following reduction of BCL-2 expression, cells become vulnerable to therapeutic agents. One method to down-regulate gene expression is antisense or gene-therapy technology. In this approach, single-stranded oligonucleotides (18-21 mer)

are designed to hybridize specifically to their region in the mRNA. Next, an RNase H complex cleaves this hybrid. A critical issue in oligo-antisense-based therapy is maintaining oligonucleotide stability in cells at the target site. To address this, researchers have introduced chemical modifications to the oligonucleotide backbone, such as replacement of a phosphate bond with phosphorothioate, base modifications, sulfur substitutions, terminal modifications and 2'-O-methyl changes in the ribose sugar. Another important problem in antisense oligonucleotide therapy is delivery of the oligonucleotide to the target site<sup>32-36</sup>.

In this study, to improve stability at the target site, we designed an antisense oligonucleotide containing phosphorothioate at both termini. To address the delivery issue, we used Lipofectamine to carry the newly designed ASO into cells, optimizing cellular uptake and endosomal escape to facilitate membrane crossing. We then investigated the effects of docetaxel and this new antisense oligonucleotide on apoptosis in LNCaP and PC-3 prostate-tumour cell lines. We designed an experiment to evaluate the efficiency of the new ASO in sensitizing cells and inducing apoptosis by down-regulating BCL-2 expres-

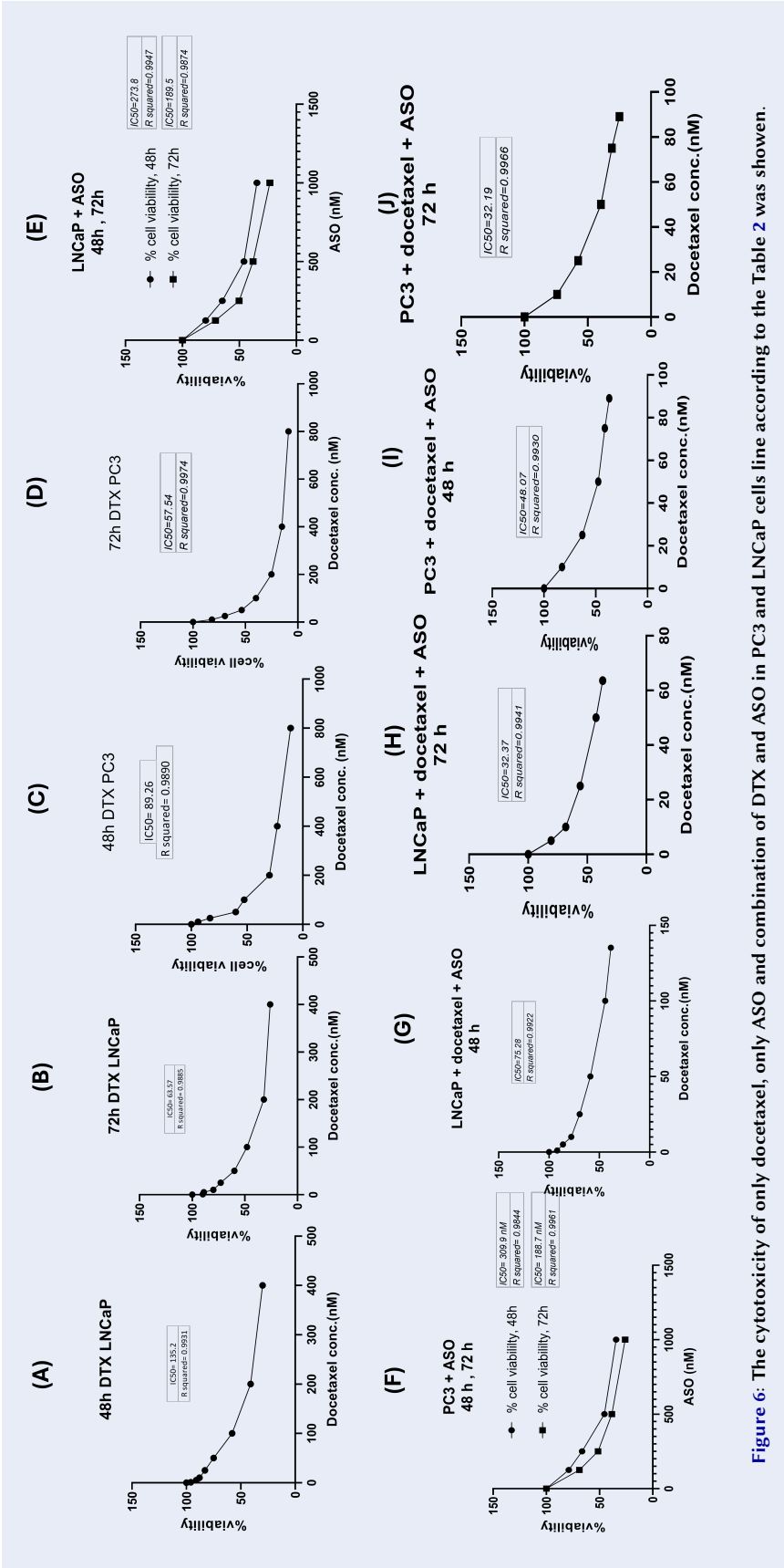
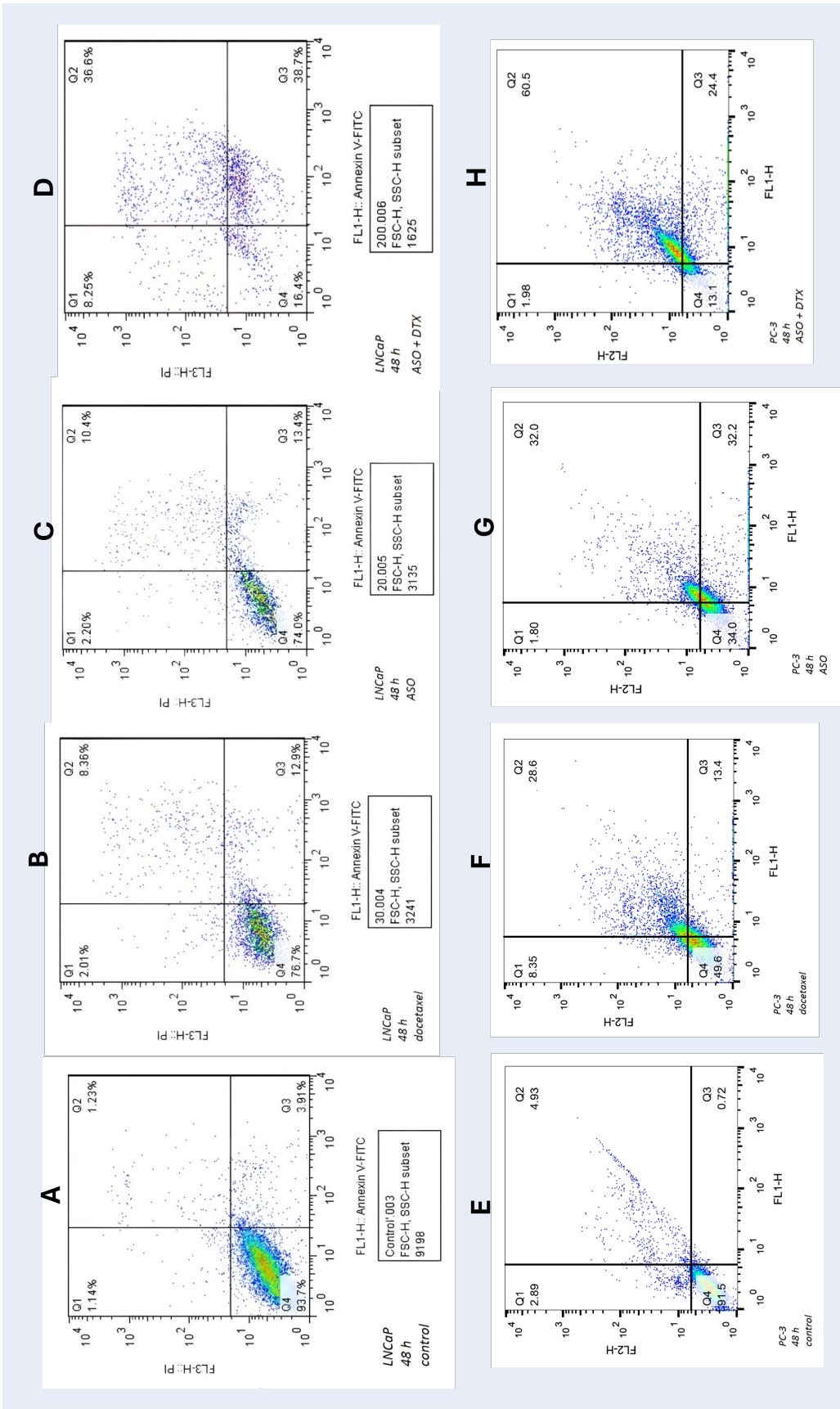


Figure 6: The cytotoxicity of only docetaxel, only ASO and combination of DTX and ASO in PC3 and LNCaP cells line according to the Table 2 was shown.



**Figure 7: The results of Flow cytometry of LNCaP cell lines.** A: The control cells, B: Treated with the new designed ASO, D: Treated with the new designed ASO and Docetaxel (DTX). The results of Flow cytometry of PC3 cell lines. E: The control cells, F: Treated with Docetaxel (DTX), G: Treated with the new designed ASO, H: Treated with the new designed ASO and Docetaxel (DTX). Cells were analyzed in independent triplicates of each combination. FL1 channel was used to detect Annexin V-FITC and FL3 indicated PI signals. The viable cells (Annexin V<sup>-</sup>/PI<sup>-</sup>) are in the Q4. Early apoptotic cells (Annexin V<sup>+</sup>/PI<sup>+</sup>) are in the Q3. Late apoptotic cells (Annexin V<sup>+</sup>/PI<sup>+</sup>) are in the Q2 and necrotic cells (Annexin V<sup>-</sup>/PI<sup>+</sup>) are in the Q1.



**Table 3:** Result of calculating synergistic effect of combination two drugs in LNCaP cell line and in PC3 cell line using bliss independence model

Cell line	ASO (%)	DTX (%)	Observed combination (%)	Bliss expected (%)	Effect
LNCaP	23.80	21.26	75.30	40.00	Synergy
	22.25	20.10	72.50	37.88	Synergy
	25.20	22.39	78.20	41.95	Synergy
	Mean ASO	Mean DTX	Mean Observed combination		
	23.75	21.25	75.33		
	SD ASO	SD DTX	SD Observed combination		
	1.48	1.15	2.85		
PC3	64.20	42.00	84.90	79.24	Synergy
	61.45	40.25	81.50	76.97	Additive
	66.95	43.75	88.30	81.41	Synergy
	Mean ASO	Mean DTX	Mean Observed combination		
	64.20	42.00	84.90		
	SD ASO	SD DTX	SD Observed combination		
	2.75	1.75	3.4		

**EBliss** = ASO + DTX – (ASO × DTX / 100)  
**ASO:** The percentage of apoptotic cells treated with ASO  
**DTX:** The percentage of apoptotic cells treated with DTX  
The percentage of apoptosis rate defined as the total of Q2 + Q3

sion. Two prostate cancer cell lines, LNCaP and PC-3, were used. Four groups of cells were prepared for each line. The first group served as control. Docetaxel was added to the second group. The new ASO was added to the third group. The combination of ASO and docetaxel was added to the fourth group. Real-time PCR was used to assess BCL-2 expression relative to the housekeeping gene GAPDH. Four ASO concentrations were tested for 48 h and 72 h. We selected the 48 h results because after 72 h flow-cytometry analysis showed necrosis in both lines. BCL-2 expression relative to GAPDH in control cells was set to 100 %. After 48 h, BCL-2 expression decreased by ~50 % compared with the control group in both LNCaP and PC-3 lines at 250 nM. Flow cytometry was used to evaluate apoptosis under three conditions: docetaxel alone, ASO alone and

the combination, all compared with control. After 48 h, 93 % of control LNCaP cells and 91 % of control PC-3 cells were alive. In groups treated with docetaxel alone, 76 % of LNCaP cells and 49 % of PC-3 cells were alive. In groups transfected with the ASO alone, 74 % of LNCaP cells and 34 % of PC-3 cells were alive. Ultimately, in the combination group, 16 % of LNCaP cells and about 13 % of PC-3 cells were alive<sup>33</sup>. Importantly, ASO off-target effects require careful consideration. A systematic approach is crucial, starting with comprehensive in-silico analysis. This should include advanced BLAST searches considering near-cognate sequences, mismatches and wobble base-pairing to predict all potential unintended binding sites across the transcriptome. Empirical validation is then necessary. Statistical rigour is es-

sential to avoid false positives. ASO design must be optimized to minimize off-target effects while maintaining efficacy. Strategies include introducing mismatches at unintended binding sites, using chemically modified ASOs for improved specificity or employing delivery methods that enhance tissue targeting and reduce systemic exposure. An iterative design–test–refine process is crucial for developing a specific and safe ASO.

This study also investigates a synergistic drug-combination strategy to enhance therapeutic effectiveness, reduce dosage and side-effects, and improve patient tolerability compared with monotherapy by accelerating cellular drug entry. In both LNCaP and PC-3 cell lines, a synergistic effect was observed. These findings offer potential for clinical trials designed to overcome DTX resistance. We recognise that our findings, while promising in established cell lines and through in-silico modelling, represent only a preliminary step. The absence of in-vivo validation limits our ability to translate these results directly to the clinic. Future studies are essential to confirm these observations in more physiologically relevant contexts. Specifically, in-vivo experiments using animal models that mimic the human disease are needed to assess efficacy and safety. Investigations using primary cells isolated directly from patients would provide a more accurate representation of disease biology and address potential limitations arising from genetic drift and adaptation of long-term cell lines. These efforts will be crucial for validating our hypotheses and paving the way for potential clinical applications.

Relying on the literature, BCL-2 up-regulation has been invariably implicated in docetaxel resistance across both castration-resistant (e.g. PC-3) and androgen-sensitive models (e.g. LNCaP). For instance, PC-3 sub-lines resistant to docetaxel exhibit elevated BCL-2/BCL-xL levels, while in LNCaP derivatives, androgen-deprivation-induced AR activation correlates with increased BCL-2 expression. Our observation that combining ASO with docetaxel in LNCaP cells lowers the IC<sub>50</sub> suggests that ASO may down-regulate this AR/BCL-2-mediated resistance axis, providing a rationale for extending this combination even to androgen-sensitive prostate-cancer models.

## CONCLUSIONS

Generally, it can be concluded that in most steps of investigation and research work, the newly designed ASO can reduce expression of the BCL-2 gene. The newly designed ASO alone was able to

reduce BCL-2 expression significantly. Flow cytometry showed that docetaxel (DTX) induced about 30-40 % of cells to enter the early stage of apoptosis, while the newly designed ASO induced approximately 30-60 %. When the combination of the newly designed ASO and docetaxel (DTX) was used, it induced 80-90 % of cells to enter early apoptosis. This makes it reasonable to conclude that by reducing expression of the BCL-2 gene, cancer cells may be more likely to undergo apoptosis sooner. A more definitive conclusion may be obtained with further studies.

However, *in-vivo* validation and primary patient-derived cells were not included in the current study. This constraint was primarily due to limited access to clinical samples, ethical considerations, and resource limitations. We used certified human prostate-cancer cell lines LNCaP and PC3 from a reliable biobank; these models are widely accepted for basic studies. We acknowledge this translational limitation and aim to address our findings in future *in-vivo* studies.

Although this study confirmed BCL-2 mRNA down-regulation, potential interactions with other BCL-2 family members such as BCL-xL or MCL-1 were not assessed. This will be examined by RNA-seq or qRT-PCR in future work for further validation.

## ABBREVIATIONS

**ASO** - Antisense, Oligonucleotide; **BCL-2** - B-cell, Lymphoma, 2; **cdNA** - Complementary, DNA; **DMSO** - Dimethyl, Sulfoxide; **DTX** - Docetaxel; **FBS** - Fetal, Bovine, Serum; **FITC** - Fluorescein, Isothiocyanate; **GAPDH** - Glyceraldehyde, 3-Phosphate, Dehydrogenase; **G3139** - Oblimersen; **IC<sub>50</sub>** - Half, Maximal, Inhibitory, Concentration; **MFE** - Minimum, Free, Energy; **mRNA** - Messenger, RNA; **MTT Assay** - (3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyltetrazolium, Bromide), Assay; **PI** - Propidium, Iodide; **PS** - Phosphorothioate; **qRT-PCR** - Quantitative, Real-Time, Polymerase, Chain, Reaction; **RNA** - Ribonucleic, Acid; **RPMI-1640** - Roswell, Park, Memorial, Institute, 1640, Medium; **SEM** - Standard, Error, of, the, Mean; **2'-MOE** - 2'-O-(2-methoxyethyl).

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## AUTHOR'S CONTRIBUTIONS

RA wrote the original manuscript provided laboratory work; MSJ analyzed the data, MM designed tables and scientific illustrations; AASM checked the

associated database and raw data and supervised; MR, RG and SS provided laboratory assistant; AM revised the final manuscript. All authors read and approved the final manuscript.

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## AVAILABILITY OF DATA AND MATERIALS

Data and materials used and/or analyzed during the current study are available from the corresponding author on reasonable request.

## ETHICS APPROVAL AND CONSENT TO PARTICIPATE

The ethics code is <http://ethics.research.ac.ir/IR.GOUMS.REC.1398.167>. This project did not involve human or animal subjects, nor did it collect data from them.

## CONSENT FOR PUBLICATION

Not applicable.

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## COMPETING INTERESTS

The authors declare that they have no competing interests.

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