Open Access Full Text Article

Chemicals used for the induction of Alzheimer's disease-like cognitive dysfunctions in rodents

Onesimus Mahdi^{1,2}, Mohamad Taufik Hidayat Baharuldin¹, Nurul Huda Mohd Nor¹, Samaila Musa Chiroma^{1,3}, Saravanan Jagadeesan^{1,4}, Mohamad Aris Mohd Moklas^{1,*}



Use your smartphone to scan this QR code and download this article

¹Department of Human Anatomy, Faculty of Medicine and Health Sciences, Universiti Putra Malaysia, Sri Serdang, 43400, Selangor, Malaysia

²Department of Human Anatomy, College of Medical Sciences, Gombe State University, Gombe 760211, Nigeria

³Department of Human Anatomy, Faculty of Basic Medical Sciences, University of Maiduguri 600230, Borno State, Nigeria

⁴Department of Human Anatomy, Faculty of Medicine and Health Sciences, Universiti Tunku Abdul Rahman, Sungai Long Campus, 43000, Kajang, Selangor, Malaysia

Correspondence

Mohamad Aris Mohd Moklas,

Department of Human Anatomy, Faculty of Medicine and Health Sciences, Universiti Putra Malaysia, Sri Serdang, 43400, Selangor, Malaysia

Email: aris@upm.edu.my

History

- Received: Mar 01, 2019
- Accepted: Oct 01, 2019
- Published: Nov 27, 2019

DOI : 10.15419/bmrat.v6i11.575

Check for updates

Copyright

© Biomedpress. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.



ABSTRACT

Alzheimer's disease (AD) is the most frequent and multifactorial form of dementia, characterised by multiple cognitive impairments and personality changes. Different methods including chemicals have been used to induce AD-like symptoms in rodent in order to screen many therapeutic drugs for a variety of cognitive dysfunctions. Articles from reliable databases such as Google Scholar, Science Direct, PubMed, Scopus, and Ovid were searched and retrieved with the following descriptors: 'Alzheimer's Disease', Cognitive impairments', Neurotoxins that induce AD', Alzheimerogenic chemicals', excitotoxins', Amyloid beta', neurofibrillary tangles. A number of chemicals have been studied to develop an animal model of AD on the basis of their mechanism of action for cognitive dysfunctions. Some of such chemicals are Heavy metals, Scopolamine, Ethanol, Colchicine, Streptozotocin, Lipopolysaccharide, and Okadaic acid among others, with a view to understanding the pathogenesis of this devastating disease. The purpose of this review is to put forward some AD pathophysiology including AD causative theories and also highlight some Alzheimerogenic chemicals for the purpose of enriching our existing knowledge. It is worth mentioning that not all the biochemical, histopathological, cognitive and behavioural abnormalities can be recapitulated. Nonetheless, experimental models of AD produced by chemicals offer insights to unravelling the pathogenesis of the disease.

Key words: Alzheimerogenic chemicals, Cognitive dysfunction, Alzheimer's disease, Alzheimer's pathogenesis, Amyloid beta

INTRODUCTION

As the aging population is increasing, the rate of age-associated diseases among older adults has become a serious health concern worldwide¹. It is wellestablished that these age-related diseases cause progressive and irreversible loss of neurons and subsequently lead to dementia. One of such age-related disorders is Alzheimer's disease (AD). AD, a neurodegenerative disease, is one of the most common and multifactorial forms of dementia characterized by multiple cognitive impairments, personality changes, and abnormal behavior². The key pathological emblems observed in AD brain tissues are amyloid beta $(A\beta)$ peptide and hyperphosphorylated tau (p-tau) protein, although the exact mechanisms which cause these alterations are yet to be uncovered³. The formation of A β is due to the aggregation of extracellular senile plaques (SP), while neurofibrillary tangles (NFTs) are caused by abnormal deposits of p-tau proteins³. Furthermore, mitochondrial dysfunction, synaptic damage, inflammatory responses, defective neurotransmissions, hormonal alterations and abnormali-

ties in the cell cycle are all linked with AD⁴⁻⁶. The remarkable risk factors linked with the progression of AD are old age, as well as multiple genetic and environmental factors. Although the etiology of AD remains unclear, multiple findings have revealed that oxidative stress is an early characteristic of the AD pathological process and also involved in the formation of A β and NFTs⁷. Thus, several attempts are ongoing in order to establish anti-AD drugs that could target specific pathogenesis, but these have not garnered much success. On the other hand, animal models have been used to trigger pathological changes similar to the human form of AD and to identify the pathogenesis, especially during the pre-symptomatic stage⁸. There is a dearth of knowledge with respect to chemically-induced AD models. Hence, this review is aimed at exploring some of the chemicals that are used for the induction of AD-like pathologies and behavioral deficits.

HISTORY OF AD

AD was first reported by Alois Alzheimer in 1906, but it was not until around 70 years after its discovery that

Cite this article : Mahdi O, Baharuldin M T H, Mohd Nor N H, Chiroma S M, Jagadeesan S, Moklas M A M. **Chemicals used for the induction of Alzheimer's disease-like cognitive dysfunctions in rodents**. *Biomed. Res. Ther.;* 6(11):3460-3484.

much interest was placed on AD research⁹. The findings that have emerged have unraveled and demonstrated, importantly, that the effects of AD commence a long time before its symptoms manifest¹⁰. Several lines of evidence have revealed that AD is a prevailing root of dementia and a significant source of death in the world^{11,12}. Since its discovery, AD has received substantial attention among the types of dementias that are of global health concern, over the years, due to its debilitating nature. To date, the precise etiology of AD remains to be elucidated, as well as questions such as why the disorder and its symptoms advance so quickly in some people (but is delayed in others), and how the disorder could be more effectively managed or treated¹⁰.

CHARACTERISTICS AND SYMPTOMS OF AD

AD is characterized by loss of memory, mood swings, problems with attention and orientation, and difficulties in carrying out daily activities. The two main pathological hallmarks of AD are aggregations of amyloid plaques (extracellular) formed by $A\beta$ and NFTs (intracellular), produced by the hyperphosphorylation of tau protein³. These alterations are ultimately followed by severe damage and loss of neurons in the brain regions concerned with memory and learning¹⁰. AD is distinguished at the cellular level by mitochondrial dysfunction, oxidative stress¹³, metal imbalance, inflammation, and apoptosis, among other hallmarks^{14,15}.

Furthermore, several symptoms are observed in individuals living with AD which changes over a period of time. Of the symptoms reported, loss of memory in recalling recent conversations are some of the early clinical manifestations; as well, depression and apathy have been identified in AD.

Additionally, other symptoms that appear later in life as the disease progresses include amnesia¹⁶, disorientation, impaired communication, confusion, poor judgement, difficulty in swallowing, speaking, and walking¹⁰. These important changes seen in AD patients reflect the severity of neuronal loss in different regions of the brain. There are three stages in AD: early, moderate and severe- through which the symptoms of the disease progress and differ from individual to individual. During the early stage of AD, patients can carry out basic things independently with little assistance for some activities; these activities include driving, walking, and other hobbies that can still be done by the patients^{10,17}. However, during the moderate stage of AD, patients may be unable to

3461

carry out routine activities, may become disoriented, and even develop personality disorders and behavioral changes (such as agitation and suspiciousness). Once it reaches the severe stage of AD, the patients become dependent on people in doing daily activities, such as eating, dressing, and bathing, among other activities¹⁷.

PREVALENCE OF AD

AD is the primary root cause of dementia; as of 2015, around 46.8 million people worldwide suffered from dementia¹⁸. This number is expected to increase exponentially to reach 131.5 million by 2050 if there are no interventions^{18,19}. In terms of prevalence between genders, AD is more of an old age disease, and women have been reported to have a longer life expectancy when compared to men²⁰. Hence, women account for about two-thirds of the elderly population affected by AD. Indeed, AD has a direct economic burden worldwide, with studies indicating that AD management cost 818 billion USD in 2016; in 2018, the expected projected cost rose to 1 trillion USD^{21,22}.

AD CAUSATIVE HYPOTHESES TAU HYPOTHESIS

The tau hypothesis suggests that hyperphosphorylation of tau protein leads to the conversion of normal tau into the paired helical filament (PHF-tau) and NFTs^{23–29}. Previously, A β had been the focus of AD research. Only recently have researchers begun to shift focus to tau protein due to the fact that a variety of reports have shown tau proteins being among the key elements contained in the NFTs. Tau protein, a member of the microtubule-associated protein, is also a functional monomeric and unfolded cell membrane protein, located within the cytosol of a neuron and very crucial in tubulin stabilization³⁰. In addition, tau is known to control neurite growth and have a role in axonal guidance, thus enhancing the normal function of neurons³¹. In humans, tau protein is only found in trace levels in non-neuronal cells³². Tau goes through various post-translational changes, especially hyperphosphorylation, a process which acts as a significant factor in influencing the stability of microtubules, thereby leading to tau protein accumulation in AD³⁰. Previous findings have shown that hyperphosphorylation of tau protein occurs via conformational changes, which are followed by the transformation of tau monomer to tau oligomer, leading to the paired helical filament and NFT formation 33-37. Recent studies have revealed that NFT itself does not appear to be implicated in causing neurotoxicity leading to the onset of neurodegeneration³⁸, though the hyperphosphorylated tau that disintegrates to form oligomers (the toxic form of tau) is implicated in neuronal damage³⁹. In addition, recent *in vivo* and *in vitro* findings in rats have highlighted that tau pathology may have far-reaching effects in distinct brain regions^{39–41}. Tau, a microtubule-associated protein, acts as a scaffolding component to assist in stabilizing microtubules by influencing tubulin stability in order to control the normal neuronal function^{42,43}.

The position of the tau gene, microtubule-associated protein tau (MAPT), is essential and located on 100 kb of the long arm of human chromosome 17 at locus 17q21, and has 16 exons⁴⁴. Moreover, it has been shown that in the brain of humans, the tau proteins encode six isoforms that have different sizes, with their length range being between 352 and 441 amino acids⁴⁵. These isoforms have some variations, three repeats (3R) or four repeats (4R) in the C terminal, and also the presence or absence of one (29 amino acids) or two inserts (58 amino acids) in the Nterminal part, which bind to actin proteins and differentiate them. Findings have revealed that the repeat areas (244-268 amino acids) located in the C-terminal are the main domain, which causes the clinging of tau to microtubule^{46,47}. Furthermore, findings have suggested that tau phosphorylation at specific epitopes could adjust the capability of tau to cling to microtubule⁴⁸. Many post-translational changes, mostly tau hyperphosphorylation, were considered to be involved as essential elements that influence the assembly of microtubule which, in turn, cause tau accumulation in AD⁴⁹. Thus, tau protein goes through conformational alterations whereas the transformation of tau monomer to tau oligomer causes the accumulation of tau, thereby causing it to pair with a helical filament and produce NFTs (due to hyperphosphorylation of tau). Emerging studies have revealed that NFTs are not associated with causing neurotoxicity. Nonetheless, the intermediary tau oligomer was reported to be a toxic form of tau that is implicated in synaptic destruction in AD⁵⁰. Tau pathophysiology is accompanied by abnormality of amyloid precursor protein (APP) and is eminently seen in the brain regions implicated in the memory-hippocampus as well as parts of the cerebral cortex⁵¹.

AMYLOID CASCADE HYPOTHESIS

Although the exact etiology of AD remains controversial, the amyloid cascade hypothesis has been widely accepted and is the most well-studied of the hypotheses out there²³. The presence of amyloid plaques is, inarguably, a key characteristic of the pathology of AD. The amyloid cascade theory proposes that amyloid plaques, made by the accumulation of $A\beta$ peptides which resulted from the proteolytic separation of APP, are crucial in AD pathology⁵². Studies have shown that the main composition of amyloid plaques in AD are polypeptides (about 4 kDa) that are usually produced in soluble form. $A\beta$ protein has been shown to have a variety of isoforms, mostly ranging from 39 to 43 amino acids^{52,53}. The two isoforms of APP, APP751, and APP770 are made up of 56 amino acids in their ectodomain.

Furthermore, findings have revealed that $A\beta$ 1-40 ($A\beta$ 40) and $A\beta$ 1-42 ($A\beta$ 42) are the two major occurring isoforms⁵⁴. Abnormal processing of APP has been found to lead to the formation of disproportionate insoluble $A\beta$ isoforms. These assemble and establish aggregates that consist of amyloid protein in the form of oligomers and protofibrils. Indeed, previous findings have suggested that this results from the changing of $A\beta$ monomers to the $A\beta$ oligomers before accumulation⁵⁵. The oligomers eventually cause damage to the neurons⁵⁶. Thus, high concentrations of indissoluble and likely $A\beta$ 42 oligomers are involved in the synaptic elimination during early stage AD⁵⁷.

Formation of $A\beta$

A β peptide^{52,58} is a derivative of Amyloid β precursor protein (A β PP), also an intrinsic type I glycoprotein⁵⁹ which has a broad ectodomain. The position of the chromosome for $A\beta PP$ is found on 21g21.2. There are 18 exons in the APP gene that exceeds 170 kb, thereby creating 10 isoforms through discrete splicing. These isoforms measure between 563 and 770 amino acids. APP is 695 amino acids in length and is one of the isoforms that has been previously reported to be found in neurons of the central nervous system⁶⁰. The specific area that codes A β -strings is made up of 16 and 17 exons, of which therein are (40 and 43) amino acid residues. Specifically, this area continues from the ectodomain to the protein's transmembrane domain⁵³. APP's central domain possesses strong affinity; hence, it can bind to ions such as copper (Cu) and Zinc (Zn), as well as heparin and collagen, within the extracellular matrix, thereby mediating the interplay of APP with the extracellular matrix. The sphere implicated in the neuritic process is defined by a precise string which is seen following the inclusion of exon 7 product⁶¹. APP isoforms have been known to play vital roles in enhancing coordination between cells and, thus, a few of those roles are hereby outlined⁶². APP isoform (of 695 amino acid) has been shown to be involved in aiding coordination between cells and enhancing the connection to the extraneuronal matrix, thereby leading to stability. APP within the intracellular domain may be associated with a cytoskeletal system that transports constituents within a particular cell. APP1-671 (β APPs) and APP1-687 (α APPs) are the other two APP isoforms produced in segments; they offer a protective role to the neurons and regulate events at the synapse⁴⁴.

Processing of APP

Proteolytic series of events taking place around and in the APP transmembrane sphere yield a better expression for the production of toxic A β 42 protein, and eventually for AD pathogenesis. Three different divisions release the APP ectodomain from the membrane. These divisions have been confirmed to be α -secretase split (which separates A β -domain and inhibits $A\beta$ -development- by generating amyloidogenic substance- due to the incapability of developing pathogenic A β)⁶³, β -secretase cleavage (the segment produced from α -secretase, while the β -secretase segments remains connected to the membrane; following further conversion by γ -secretase it becomes weakened), and A β -protein (A β 42) breakage (whose aggregation is thought to cause AD due to the presence of toxic and fibril aggregates) 52,54,58.

Biochemistry of Senile Plaques

Blocq and Marnesco, in 1892, were the first to demonstrate that senile plaques (SP) could be described as densely packed structures called amyloid bodies. The process by which amyloid bodies develop and are implicated in AD development is known as amyloidosis. Fibrillogenic proteins (about 10 nm in diameter; amyloid fibrils that are smooth and straight) and non-fibril components (ApoE and serum A β components) are the contents of the amyloid bodies. The SPs, which are also called amyloid plaques, are the product of an extracellular accumulation of A β protein ⁶⁰.

CHOLINERGIC HYPOTHESIS

The oldest theory among the AD causative theories is the cholinergic hypothesis⁶⁴. This hypothesis states that a decrease in neurotransmitters, known as acetylcholine, in neurons is responsible for AD etiology. The cholinergic hypothesis has been postulated for more than 3 decades now and suggests that abnormal acetylcholine-containing neurons in the basal ganglia are implicated in cognitive decline seen in AD patients⁶⁵. Essentially, acetylcholine (ACh) is a neurotransmitter utilized by cholinergic neurons and is crucial for physiological processes like attention, learning, memory, stress response, wakefulness, and sleep, as well as sensory information^{66–70}. Damage to cholinergic neurons was observed as an important pathological alteration which corresponds with cognitive destruction seen in AD.

Consequently, this hypothesis was initially tested using cholinesterase inhibitors which are used for AD treatment. Thus, as a result of the trial, it has been noted that tacrine, one many cholinesterase inhibitors (including donepezil, galantamine, rivastigmine, and memantine), was the earliest anti-AD drug to be used in the clinic 71,72. However, due to some identified severe side effects, it has been retrieved from the market since 2012. Even though cholinesterase blockage is a typical palliative treatment with minor gain, presently, it appears to be the only obtainable clinical remedy that offers hope to despairing patients with AD. Despite these facts, the cholinergic hypothesis has not been widely accepted, mainly due to the fact that the medication was proposed to treat deficiency of acetylcholine but was not effective. Nonetheless, about 4 of the 5 approved anti-AD drugs available in the market today were developed based on the cholinergic hypothesis²³.

OXIDATIVE STRESS HYPOTHESIS

Oxidative stress is believed to be critical in AD pathogenesis⁷³. Notably, the brain is known to consume more energy and exert more functions than any other organs during mitochondrial respiration, which also increases the likelihood of reactive oxygen species (ROS) exposure 74. Furthermore, AD is closely linked to molecular oxidative stress, as well as protein nitration, the rise of protein oxidation, lipid peroxidation, and glycoxidation. Moreover, AD is closely linked to the aggregation of A β , which has been reported to cause oxidative stress⁷⁵⁻⁸⁰. Hence, antioxidant compounds are good candidates to offer protection against oxidative stress and A β toxicity, to a certain extent. Although oxidative stress is one aspect of AD, the antioxidant approach has been disputed, too, since it lacks the potential to impede the advancement of AD, and has been suggested to be used as part of combination therapy^{81,82}.

INFLAMMATION HYPOTHESIS

Recent studies have uncovered that neuroinflammation and aberrant gliosis are also emblems of AD⁸³. Indeed, the inflammation hypothesis has been validated by genetic and transcriptomic studies^{84–87}. Microglia-related pathways were observed as crucial risk factors for AD and its pathogenesis. A wealth of information has indicated that microglia is a crucial factor. For instance, during the initial phase of AD, microglia and its associated proteins, such as the Triggering Receptor Expressed on Myeloid Cells-2 (TREM2), can influence synaptic reduction^{88,89}. The process of activity-dependent and long-term synaptic plasticity is the typical and intrinsic molecular basis of learning and memory that could be the observable effects on long-term potentiation⁹⁰. Thus, subsequently, amyloid plaques will be surrounded by aberrant microglia and astrocytes, and produce several proinflammatory cytokines. These series of events are the steps involved in the evolution of AD. However, in the clinic, non-steroidal anti-inflammatory drugs (NSAIDS) have not appeared to be beneficial. This could be largely due to the link between innate immunity and the complexity of AD pathogenesis; thus, the immune responses generated could either be detrimental or useful based on the context^{87,91,92}.

Nonetheless, there are new findings that indicate that the PD-1 immune crossing point barrier decreases the pathology of AD, thereby enhancing memory in AD mouse models^{93–95}, and has become an area for subsequent research. The recent awareness towards uncovering the mechanisms involved regarding microglia disruption in clipping, neurogenesis and plasticity regulation are unlocking areas, allowing for the exploration of better therapeutic interventions and diagnoses of AD^{96,97}. Understanding the abnormal microglial roles and restoring homeostasis could provide a new set of concepts for treating AD. However, due to the intricacy and distinct roles of microglia in health and disease, new biomarkers that reflect the functions of specific microglia are critically needed 92,98.

Despite all the above armamentarium, AD still needs to be properly understood in order to address its remote causes and the mechanisms underlying its progression, of which chemicals or drugs have been indicated as some of the causative factors.

MITOCHONDRIAL CASCADE HYPOTHESIS

Swerdlow was the first person to propose the mitochondrial cascade theory in 2004, where he posits that mitochondrial abnormality is the main reason for A β accumulation, the formation of NFT, and degeneration of synapses in AD²⁴. Mitochondrial causative theory utilizes many theoretical liberties. It presumes that the physiologic mechanisms underlying AD and the aging brain are similar. It states that based on the fact that mitochondrial dysfunction in AD is systematic, mitochondrial dysfunction is not enough to represent the effects of neuronal degeneration.

Furthermore, the hypothesis of the mitochondrial cascade forecasts that non-Mendelian inheritance is linked to non-autosomal dominant AD²³. Lastly, it postulates that AD brain mitochondrial abnormality propels amyloidosis, phosphorylation of tau, and cell cycle re-entry. Mitochondrial dysfunction is detected in many AD tissues²⁵, which include fibroblasts, platelets, mitochondria, and brain. There are three main defective mitochondrial enzymes involved: α ketoglutarate dehydrogenase complex, pyruvate dehydrogenase, and cytochrome oxidase²⁶. AD brain investigations have revealed a normal concentration of cytochrome oxidase but with an altered structure of the enzymes itself²⁷. Oxidative stress and proteasome abnormality have been hypothesised to enhance mitochondrial dysfunction²⁸. Moreover, studies involving cytoplasmic hybrids (cybrid) have revealed that mitochondrial DNA (mtDNA) is partly responsible for the decreased cytochrome oxidase in AD²⁹.

COMMONLY USED CHEMICALS TO INDUCE AD-LIKE SYMPTOMS IN RODENTS

Some of the common chemicals used for modelling AD are scopolamine, streptozotocin, and alcohol, as well as dysregulation of heavy metals, such as aluminum (Al), copper (Cu), zinc (Zn), lead (Pb, and reducing sugar (D-galactose)⁹⁹, among others (**Tables 1, 2 and 3**).

Aluminum as one of the frequently used heavy metals for induction of cognitive impairment

Environmental heavy metals are agents that have been well-established to impact the development of the brain through neurotoxicity¹⁰⁰. Among the commonly used Alzheimerogenic chemicals, heavy metals have been identified to induce high levels of toxicity that is linked with several diseases, including neurodegenerative disorders, following long-term exposure in humans or chronic administration in rodents. A growing body of support has reported the relation between heavy metals and neurodegenerative diseases, including AD and Parkinson's disease ¹⁰⁰. Heavy metals are generally known to damage the nervous system. Of all the heavy metals, the effect of aluminum on biological systems has been well-reported^{45,101,102}. Al is an ubiquitously scattered environmental and industrial toxicant associated with anemia¹⁰³, osteomalacia, and hepatic and



Figure 1: Some Alzheimerogenic chemicals. The neurotoxic effects of these chemicals lead to gradual neuronal loss and thereby resulting in neurodegeneration and its associated symptoms.

neurological disorders¹⁰³. The high amount of Al has been identified in the brain of subjects suffering from AD and causes toxicological effects, including encephalopathy, bone disease, and anemia. Very recently, orally administered Al (300 mg/kg body weight) was reported to induce oxidative stress, cholinergic deficit, and accumulation of A β & NFTs in the brain of rats¹⁰¹.

Moreover, in the Al-treated group, there are marked histopathological changes and diffuse gliosis accompanied by pericellular edema in the cerebral region, in addition to neuronophagia and loss of neurons. A number of studies have shown that Al neurotoxicity is highly linked with cognitive impairment of AD, via oxidative stress and cholinergic dysfunction ¹⁰⁴. The findings have indicated an upregulation in the expression and activity of acetylcholinesterase (AChE) and malondialdehyde (MDA), but a significant decrease in expression and activity of glutathione-s-transferase (GST), glutathione peroxides (GPX), and glutathione reductase (GR).

Scopolamine-induced AD-like dementia

A growing body of knowledge has established that scopolamine is an anticholinergic drug mostly employed as an approved chemical in pilot studies to cause memory deficit¹⁰⁵. Administration of scopolamine leads to deficits in visual recognition memory, verbal recall, visuospatial praxis, visuospatial recall psychomotor speed, and visuoperceptual processes¹⁰⁶. It is one of the most potent and commonly

used drugs to prevent motion sickness¹⁰⁷. Furthermore, researchers have documented that scopolamine non-selectively occludes the adhesion site of ACh muscarinic receptors in the cerebral cortex and results in the unequal discharge of ACh, which annihilates hippocampal neurons and induces learning and memory impairment in mice in a dose-related manner^{108,109}. However, the main effects of scopolamine likely result from blockage of some receptors (M1 and M5) due to their distinct distribution in the brain¹¹⁰. Previous studies have also suggested that it may lead to memory and learning deficit in a dose-dependent way^{108,109}. In some instances, ordinary doses of scopolamine led to agitation, confusion, hallucination, paranoid behaviors, delusions, and incoherent speech 111. Previous studies have indicated there is potential participation of N-methyl-Daspartate (NMDA) receptor mechanisms of the dorsal hippocampus in memory loss due to scopolamine induction¹¹². Currently, histone deacetylase-2 and DNA methyl transferase-1 are important for synaptic plasticity induced by scopolamine after a decrease in memory¹¹³. Hippocampal administration of scopolamine occludes long-term potentiation 114 and impedes spatial encoding¹¹⁵.

Furthermore, the injection of scopolamine into the medial septum impedes learning and decreases ACh discharge into the hippocampus¹¹⁶, injection of scopolamine in the CA3 area of the hippocampus induces encoding impairments selectively, but memory retrieval in Hebb-William maze was not affected¹¹⁷.

Indeed, scopolamine has been identified to cause spatial learning and memory deficiencies. Consequently, it leads to the excitation of glycogen synthase kinase-3 beta (GSK- 3β), inadequate spine maturation, and the arborization of dendrites connected with alterations in CREC, Homer1, and amino-3-hydroxy-5-methyl-4-isoxazole propionic acid receptor (AMPA)¹¹⁸.

Colchicine-induced AD-like symptoms

Advances in chemicals used for modelling AD have suggested that colchicines are potential drugs that induce dementia through dysfunctional cholinergic neurons by either inhibiting cholinergic reversal or eroding cholinergic passage cascades^{119,120}. Colchicine induces hippocampal lesions resulting in cognitive impairments and ChAT reduction, suggesting that it can be utilized as a candidate for modelling AD. Colchicine could cause neurotoxicity and memory decline by inhibiting cholinergic pathways, thereby leading to a reduction in the number of cholinergic neurons and subsequently decreasing cholinergic renewal primarily within the hippocampal area of the brain¹²¹. Memory impairment seen induced by colchicine may be a result of a decrease in serotonin, dopamine, and norepinephrine within the caudate nucleus, hippocampus, and entire cerebral cortex¹²².

Furthermore, colchicine was shown to cause the production of protein carbonyls following lipid peroxidation¹²³. Colchicine has also been found to raise cyclooxygenase-1, and 2 (COX-1 and COX-2) expression levels¹²⁴ and ROS generation¹²⁵. Colchicine raises glutamate/GABA ratio in the cortex¹²⁶ and triggers excitation of MDA receptors that lead to a sharp rise in the influx of Ca^{2+} , therefore resulting in the triggering of enzymes that depend on Ca²⁺ (e.g. phospholipases A2, xanthine oxidase, proteases, cyclooxygenases, and protein kinases) 127. Intracerebroventricular (ICV) administration of colchicine (7.5 g in 10 L) was found to recapitulate cognitive memory decline in rats¹²⁸ and similarly in mice¹¹⁹. A significant memory deficit was observed two weeks after cognitive induction impairment using colchicine¹¹⁹. Furthermore, 3 g/mice (ICV injection) of colchicine induces spatial memory impairment¹²⁹. The main advantage of this model is the fact that it causes some symptoms of sporadic Alzheimer's type of dementia, similar to those found seen in human subjects, like time-variant changes in onset, behavioral, and biological patterns¹²³.

Streptozotocin-induced AD-like cognitive decline

Streptozotocin (STZ), a compound derived from glucosamine nitrosourea and found in the strain of Streptomyces achromogenes, when administered ICV in rodents¹³⁰, is known to exert a severe and long-lasting effect on the brain's cytoarchitecture, biochemistry, metabolism, and functions (such as decreased glucose uptake and energy consumption, oxidative tissue stress, cholinergic differential, and cognitive abilities). STZ is another alkylating agent used in cancer treatment. It has also been found to mimic certain properties of nitrosourea, an anti-cancer agent that has been reported to have hypoglycemic effects in addition to being involved in memory damage¹¹⁰. Induction of neuronal damage and hyperphosphorylation of tau is caused by STZ, resulting in the release of ROS and reactive nitrogen species (RNS)¹³¹. Findings revealed that STZ destructs the glycolytic enzyme activity within the brain which eventually results in the lowering of ATP and creatine phosphate concentrations. This destructed energy system and lowered acetyl CoA synthesis can inhibit cholinergic conductance¹³⁰. Rats induced by STZ have been reported to demonstrate raised activities of AChE in their brains and decreased ACh. In other studies, STZ has been found to activate A β peptide-like aggregates by modifying the GSK alpha/beta¹³².

Furthermore, there is evidence that gene expression associated with the development of glial-derived NF, brain-derived neurotrophic factors (BDNF) and integrin -alpha-M becomes increased by STZ, while expression of genes for NGE-IB and metallothionein 1/2 is down-regulated and eventually results in the modification in apoptosis and cell survival process¹³³. Administration of ICV-STZ resulted in a substantial decrease in memory and learning skills of rats, leading to oxidative stress¹³⁴. Additionally, ICV administration of STZ (3 mg/kg) in a model of cognitive impairment in rats mimicked the sporadic dementia of AD¹³⁵. Memory impairment was observed in the rats during the last phase of the experiment using the Morris water maze (MWM) task in order to assess AD-like symptoms. In addition, impaired glucose metabolism, a decrease in cholinergic markers, and oxidative stress were seen in the rats' brains. Altogether, these long-lasting effects mimicked those seen in AD patients.

Table	Table 1: Studies showing Alzheimerogenic Chemicals									
S/N	Species	Chemical	Age/weight/rou of administratio	Behaviour	Biochemical Assay	Histopathology	Ref			
1.1	Rats	Al	150 – 170 g 3 times a week 50 mg/kg 90 days Orally	Short-term & long term cognitive disturbance Slow locomotor activity	↓ AChE ↓ CAT, GSH	-Neuronal loss -Vacoulated cytoplasm	136			
1.2	Female Sprague- Dawley Rat	Al. Cu & Fl Orally	170 – 200 g 50, 5 & 20 mg/kg 40 days	Significant learning deficit	$\downarrow GSH, SOD \downarrow GST & GPx \uparrow proinflam-matory TNF-\alphaIL-1\beta & IL-12↑ AChE↑ APP geneexpression& A\beta42 levelA\beta accumula-tion,oxidative stress$	-Degeneration of pyra-midal neurons with Pyknotic nuclei -Swollen neurons & vacoulation Apoptotic cells -Neuronal degeneration with eisonophilic accumulation	137			
1.3	Rats	Rats AlCl ₃ Orally	180-200 g 300 mg/kg 60 days	Memory impairment Spatial Learn- ing & memory	 ↑ AChE activity ↑ proinflam- matory cytokines TNF-α, IL-1 & IL-6 ↑ BDNF mRNA levels ↓ CAT, GSH, GST & MDA levels in hippocampus 	Neuronal loss pyknosis in CA1 & CA3	2			
1.4	Rats	Rats AlCl ₃ Orally	250-300 g 17 mg/kg 21 days 12—15 weeks old	Impaired cognitive function Decrease in time to reach food & deteriorated memory	$\uparrow IL-6$ neuro-toxicity due to NO & ROS $\downarrow AChE$ activity $\downarrow ACh$ $\downarrow BDNF$ $\downarrow DA$ $\downarrow TAC$ $A\beta$ plaques formation	-↓ in cells in granular -& pukinje layers -Thin irregular reduction in cell -cell size of molecular layer -Sparsely distributed cells -Neurodegeneration in cortex -DNA damage	138			

Table 1: Studies showing Alzheimerogenic Chemicals

AChE: acetyl cholinesterase, ACh: acetylcholine, ChAT: cholineacetyltransferase, MDA: Melondialdehyde, SOD: Superoxide dismutase, iNOS: inducible nitric oxide, GSH: glutathione, GPx: glutathione peroxidase, CAT: catalase, DA: dopamine, BDNF: brain derived neurotrophic factors, NO: nitric oxide, ROS: reactive oxygen species, TAC: total antioxidant content, OG: oral gavage, ICV: intracerebroventricular, IH: intrahippocampal, SC: subcutaneous, SCM: Scopolamine, STZ: Streptozotocin, OKA: Okadaic acid, CLC: Colchicine, SD: Sprague-Dawley, IP: intraperitoneal, \uparrow : increase, \downarrow : decrease, \leftrightarrow : unchaged, LPS: Lippolysacharide, TNF- α : Tumor necrosis factor, NFTs: neurofibrillary tangles, COX-2: Cytocrome oxidase, TBARS: Thiobarbituricacid reactive substance, MPO: Myeloperoxidase, GST: Glutathione S-transferase, GFAP: Glial fibrillary acid protein

S/N	Species	Chemical	Age/weight/route of administration	Behaviour	Biochemical Assay	Histopathology	Ref
1.5	Rats	AlCl ₃ + D-gal OG & IP	280-300 g 10 weeks 300 mg/kg 60 mg/kg	Memory and Learning deficits Spent short time in the target quadrant	↑ AChE activity ↑ Protein expression	Neurodegeneration in hippocampal neurons Amyloid-like deposit	103
1.6	Rats	ICV/STZ	280-300 g 0.5 mg/kg (3-5 weeks) 1 mg/kg (9-11weeks) 3 mg/kg	Severe impairment in working memory	↓ ChAT mRNA ↓ IR expression	Astrogliosis dark neurons indicating neu- rodegeneration ↓ Astrocytes in DG, CA1 & neuronal death	139
1.7	Rats	ICV/STZ	350-400 g 2 mg/kg 3-4 months	Cognitive impairment	↓ Synapto- physin	Neuronal loss Ventricular enlargement p-tau & Aβ peptide accumulation	140
1.8	Mice	ICV/STZ Single dose	20-25 g 2 weeks	Learning & memory perfor- mance ↓ in mean time spent in a quadrant by MWM Deficit in spatial learning impairment in novelty in seeking behaviour	\downarrow α-secretase activities \downarrow cerebral Aβ42, β- secretase & COX-2	Aβ deposits	141
1.9	Rats	ICV/STZ	220-25 g 5 μl 3 months	Deficits in spatial learning & memory in MWM passive avoidance task	† Oxidative stress ↓ GSH † MDA levels	$A\beta$ deposits Shrunken pyramidal cell layer & organisation Cellular infiltration & congestion Apoptotic cells & neuronal loss	142

Table 2: Studies showing Alzheimerogenic Chemicals (Continued)

AChE: acetyl cholinesterase, ACh: acetylcholine, ChAT: cholineacetyltransferase, MDA: Melondialdehyde, SOD: Superoxide dismutase, iNOS: inducible nitric oxide, GSH: glutathione, GPx: glutathione peroxidase, CAT: catalase, DA: dopamine, BDNF: brain derived neurotrophic factors, NO: nitric oxide, ROS: reactive oxygen species, TAC: total antioxidant content, OG: oral gavage, ICV: intracerebroventricular, IH: intrahippocampal, SC: subcutaneous, SCM: Scopolamine, STZ: Streptozotocin, OKA: Okadaic acid, CLC: Colchicine, SD: Sprague-Dawley, IP: intraperitoneal, \uparrow : increase, \downarrow : decrease, \leftrightarrow : unchaged, LPS: Lippolyssacharide, TNF- α : Tumor necrosis factor, NFTs: neurofibrillary tangles, COX-2: Cytocrome oxidase, TBARS: Thiobarbituricacid reactive substance, MPO: Myeloperoxidase, GST: Glutathione S-transferase, GFAP: Glial fibrillary acid protein

Biomedical Research and Therapy, 6(11):3460-3484

S/N	Specie	sChemical		Age/weight/route Behaviour		Histopathology	Re
			of administration	on	Assay		
1.10	Rats	ICV/STZ	300-340 g 3-4 months 30 days	 Memory impairment Decrease in time spent In closed arm 0/OFF Deficits in short-term Spatial memory seen is decrease time spent on new arm Using Y Maze Short-term recognition 	Neuro inflammation	↓ Cell propagation ↓ in proliferation of marker Ki-6 & immature neurons DCX in SVZ	143
1.11	Rats	IP/SCM	150- 250 g 20 g/kg	 Memory deficit ↑ conditional avoidance 	 ↑ MDA ↑ Lipid peroxidation ↓ GSH level ↑ AChE activity 	Degeneration of neurons with pyknotic & condensed nuclear Gliosis	144
1.12	Rats	IP/SCM	200-220 g 1 mg/kg	 Severe memory impairment (ST & LT in hippocampus) Delayed latency in MWM & frontal dependent memory Task 	Occluded cholinergic signals	Altered cortico -hippocampal neurons Retraction process in pyramidal cell Layer Vacoulation of surrounding neutrophils of pyramidal cells & hyperchromatic & shrunken perikaryo Cork-screw shaped apical dendrites	145

Table 3: Studies showing Alzheimerogenic Chemicals (Continued)

				Table 3 continued			
S/N	Species	Chemical	Age/weight/rout of administration	Behaviour	Biochemical Assay	Histopathology	Ref
1.13	Rats	OG/SCM	150-200 g 2.5 mg/kg 1 hour	-Memory impairment	↑ AChE activity ↓ GABA &GSH	Neuronal degeneration Hippocampal Phalomalacia & Oedema in tissue matrix with de-myelination Congestion of blood Capillaries Perivascular Oedema in cortex	146
1.14		IP/SCM	1 mg/kg 8 weeks old	-Impairment of learn-ing & memory -Impairment of acquisition of ST & LT	↓ mRNA expression ↔ AChE	-	147
1.15	Mice	IP/SCM	10 mg/kg 7- 12 weeks	-Impaired memory -↓ ambularv movement -Deficits in social recognition	↑ Cholinergic neurones	-	148
1.16	Rats /Mice	ICV/ CLC		- Memory loss	↑ TNF-α ↑ ROS, COX2 Nitrite	-	149
1.17	Rats	ICV /CLC	15 μ1/5 μl aCSF	Significant memory loss Learning & memory deficits	↓ GSH, SOD, GST ↑ MDA levels Oxidative damage ↑ AChE activity	Destruction of neurons	150
1.18	Rats	ICV /CLC	15 μ1/15 μl aCSF	Cognitive impairment Learning and learning deterioration	↑ MDA level ↔ CAT ↓GSH ↓ AChE	Neuronal loss	151
1.19	Rats	ICV /CLC	180-200 g 15 μ1/rat 3 weeks	Memory impairment Impaired acquisition of spatial navigation task	↓GSH, Oxidative stress ↑ ROS Aβ pep- tide deposit in hippocampus	Neuronal damage \downarrow hippocampal tissue levels of BDNF $A\beta$ deposits in hippocampus Continued on next	152

Science & Technology Development Journal – Economics - Law and Management, 6(11):3460-3484

				Table 3 continued			
S/N	Species	Chemical	Age/weight/roo of administration	Behaviour	Biochemical Assay	Histopathology	Ref
1.20	Rats/ Mice	ICV/	200-250 g 20-30 g	Memory impairment	↑ TNF-α, IL-113	Neurodegenera	153
		CLC	7.5 μl in 2.5 μl 6-8 weeks		↑ ROS & nitrite	Plaque formation	
			old			Reduction	
						in the intensity of Nissl granules	
	Rats	IH/OKA	200-320 g	Decrease in time find-	↓ Glu syn-	Gliosis	154
1.21			90 days	ing platform	thetase	Astrogliosis	
			100 ng	Spatial cognitive	\downarrow GSHOxidative	\downarrow GFAP	
			12 days		stress	expression	
	Rats	ICV	300-380 g	Spatial cognitive	↓GSH	Gliosis	155
1.22	2	/OKA	100-200 ng	deficit Significant ↓ in time to find	Oxidative	p-tau-like	
					Stress Tau	formation	
				platform	phosphory- lation site at		
					396		
	SD	ICV	220-250 g	Memory deficit	↑ MDA &	Loss of	156
1.23	8 Rats	/OKA	200 ng	Poor memory perfor-	Nitrite \downarrow	pyramidal	
			13 days	mance	GSH & Lipid	Blabbing	
					peroxidation	of cells in	
						the brain &	
						sponginess	
						Synaptic	
			20.22			dysfunction	157
1.24	Mice	ICV /OKA	20-22 g 200 ng		↓ GPx ↑ MDA	Neuronal	107
1.24	ŧ	/OKA	200 lig 2 times		mitochondrial	damage Nuclear pylypooie	
			3 days		cells	Nuclear pyknosis Reduced	
			interval			density of Nissl bodies	
						Neurofibrillary	
			10.00	Cognitive	Δ TNF - 0-11 <i>Q</i>	degeneration Neuronal loss	158
1.25	Mice	OG/ Ethanol	18-22 g 12 mL/g	impairments	↑ TNF-a &IL-ß		100
			Once daily 1 st week	Short distance	↓ Glu & GABA Neurotrans-	↓ in microglial cells	
			Twice daily after 1 st	covered during spontaneous	mitter	↓hippocampal	
			week	movement	imbalances	DG cells	
	SD	OG/	257-300 g	Learning & memory	\uparrow TNF- α in	Shorter	159
1.26	6 Rats	Ethanol	5 g/kg	deficits	Hippocampus	microglia	
			Days 2 & 4		\leftrightarrow IL-10 BDNF	Lack of ED-1	
						positive cells	
						Continued on next	page

				Table 3 continued			
S/N	Species	Chemical	Age/weight/ro	ute Behaviour	Biochemical	Histopathology	Ref
			of administration		Assay		
1.27	Rats	OG/ Ethanol	180-200 g 396-426 g (after long exposure)	Memory impairment	↑ IL-15 gene NSD alter long term exposure	Reduction in the microglia shortening & of processes with brush appearance	160
1.28	SD Rats	OG/ Ethanol 4.5 mg/kg	200-300 g 21 days		↑ AChE activity	Neuronal death Apoptotic cells Neurodegen- eration	161
1.29	Rats	OG/ Ethanol 4.5 g/kg	170-220 g 21 days	Cognitive impairment ↓ discrimination in- dex in novels object discrimination	↓ AChE activity Oxidative stress	-	162
1.30	SD Rats	ICV/LPS 2 ul/1min	200-220 g 21 days	-Loss of spatial memory -Reduction in sniffing times -Less platform-cross number	↑ IL-IB in hippocampus ↑ TNF-e & COX-2 ↑ NF-Kb, iNOS	-	163
1.31	Mice	IH/LPS 40 ug/mouse single admin	18-22 g 7 days	-Learning & memory impairment -Treated mice took longer time to find platform using -MWM -Spontaneous alter-ation in Y-Maze	mRNA ↓ TNT-α, NO & IL-6 Activation of microglia in hippocampus CA1 & DG	Reduction in NeuN stained area of neurons in hippocampus ↓ in number of DCX positive cells	164
1.32	ICR Mice	IP/LPS 250 ul/kg 7 times Daily	-	Memory deficit Spent longer time in a target quadrant	\downarrow IL-1 β , IL-6 & TNF- α \downarrow GSH/GSSG \downarrow COX-2, iNOS \downarrow MDA content	Brown coloured labelled $(A\beta 42)$ cells in the hippocampus Higher GFAP Iban1- reactive cells	165
1.33	SD Rats	lP/LPS 10 mg/kg Single dose	250 g 7 —9 days		↑ TNF-α IL-1ß & IL-6	Deposits of $A\beta$ plaques P—tau inclusions in the brain	166

				Table 3 continued			
S/N	Species	Chemical	Age/weight/rou		Biochemical	Histopathology	Ref
			of administration	Behaviour on	Assay		
1.34	ICR Rats	IP/LPS 0.25 mg/kg		- Deficits in spatial memory - Decrease in latency	\uparrow Aβ342 in cortex & hippocampus \uparrow β -secretase activity \uparrow γ-secretase in Hippocampus & Cortex \uparrow iNOS, COX- 2 & GFAP	Gliosis in cortex & hippo evidence by thick & short processes	167
1.35	SD Rats	SC/NAN 4—51 mg/kg/day (mini pumps)	31 days	Learning & memory deficits Weaker cognitive per- formance	↓ ChAT & AChE activity	Pycnotic nerve cells Nerve cell loss Corkscrew- like dendrites Positively stained gran-ules by AT8-positive (tau) granule	168
1.36	Rats	IP/AN3 12.5 mg/kg/ days 10 mg/kg/ day	5 days 9 days	-Impairment of learning & memory -Spent more time explore target -quadrant	↓ GSH levels ↑ AChE, Nitrite ↑ TBARS, MPO Cytochrome C NO	Neuronal loss	169
1.37	SD Rats	SC/ NAN3	400-425 g 1 mg/kg/h 7 days	-Impairment of learn-ing & memory -Spent more time explore target -quadrant	$\downarrow Cytochrorme C \leftrightarrow NO$	-	170

AChE: acetyl cholinesterase, ACh: acetylcholine, ChAT: cholineacetyltransferase, MDA: Melondialdehyde, SOD: Superoxide dismutase, iNOS: inducible nitric oxide, GSH: glutathione, GPx: glutathione peroxidase, CAT: catalase, DA: dopamine, BDNF: brain derived neurotrophic factors, NO: nitric oxide, ROS: reactive oxygen species, TAC: total antioxidant content, OG: oral gavage, ICV: intracerebroventricular, IH: intrahippocampal, SC: subcutaneous, SCM: Scopolamine, STZ: Streptozotocin, OKA: Okadaic acid, CLC: Colchicine, SD: Sprague-Dawley, IP: intraperitoneal, \uparrow : increase, \downarrow : decrease, \leftrightarrow : unchaged, LPS: Lippolyssacharide, TNF- α : Tumor necrosis factor, NFTs: neurofibrillary tangles, COX-2: Cytocrome oxidase, TBARS: Thiobarbituricacid reactive substance, MPO: Myeloperoxidase, GST: Glutathione S-transferase, GFAP: Glial fibrillary acid protein

Ethanol/Alcohol induced AD-like symptoms

Chronic intake of alcohol is associated with many problems including attention deficits, impairment in language and social skills, hyperactivity, motor dysfunction, and learning deficits¹⁷¹. Previous studies have shown that ethanol consumption enhances the generation of ROS and results in a decrease of the antioxidants in the brain¹⁷². Another finding has suggested that ethanol could damage hippocampal and cholinergic neurons with a resultant effect on the sensory-motor system as well as disruption of learning and memory¹⁷³. High ethanol treatment resulted in excessive nitric oxide (NO), which has been found to destruct memory and learning, while higher doses of ethanol disrupted the glutamatergic system and increased GABAergic conveyance in the brain areas that are associated with memory¹⁷³. Ethanol also elevates the level of adenosine, which may, in turn, lead to memory damage¹⁷⁴ and can result in a shorter route by accelerating memory impairment and promoting active, spontaneous motion.

Furthermore, results obtained following the MWM test in mice showed a notable increase in escape latency and total swimming distance, with a sharp drop in cross time. Thus, chronic alcohol administration can significantly affect spatial learning ability and cognition by mice¹⁷⁵. However, memory impairment caused by prolonged alcohol intake can be observed from the anti-inflammatory activity and from the control of the equilibrium of Glu and GABA¹⁷⁶. The particular molecular mechanisms still need further investigations, but overall, alcohol can significantly disrupt learning and memory.

Memory deficit induced by excitotoxin

Ibotenic acid is a strong neurotoxin that aggravates signs and pathophysiology analogous to AD^{177,178}. It is a useful model to appreciate drug efficiency in evading AD pathology. It has also been found that intrahippocampal administration of ibotenic acid (5 μ g/ μ l PBS) produces memory impairment and results in increased AChE activity as well as elevated MDA levels, thereby inducing neuronal toxicity¹⁷⁹. Bilateral ICV injection of ibotenic acid may elicit AD-like pathology and symptoms. Indeed, a particular study found that ibotenic acid decreased the activity of cholinergic neurons in rats¹⁸⁰. Many other cholinotoxins and neurotoxins that present AD-like symptoms are kainic acid, quinolinic acid, anti-NGF, NMDA antagonist dizocilpine, and α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA). Previous studies

have reported that intrahippocampal kainic acid (0.4 $\mu g/2\mu l$) administration led to oxidative damage (revealed by the increase of hippocampal lipid peroxidation, nitrite level, and diminished superoxide dismutase (SOD)). Thus, it is clear that kainic acid activates oxidative damage and memory loss in rats¹⁸¹. Stimulation of glutamate receptors, particularly NMDA receptors, in a diseased condition can lead to disrupted ion homeostasis, decrease in energy, neuronal loss, and cell death (excitotoxicity) by an abnormally high concentration of glutamate^{182,183}.

Sodium azide-induced AD-like memory impairment

Sodium azide (NaN3) is a white crystalline solid that has been reported to be a mitochondrial toxin implicated in the production of lead azide explosive¹⁸⁴. NaN3 administration has been known to induce mitochondrial dysfunction and inhibit cytochrome oxidase, a critical mitochondrial enzyme¹⁶⁹. Cytochrome oxidase is critical in the mitochondrial respiratory chain; its interference impedes with mitochondrial complex-IV and decreases ATP levels, which add to metabolic impairment and ROS generation¹⁷⁰, ultimately producing some sequelae of AD. Neuronal loss in the CA3 area of the hippocampus is one of the essential features used to confirm the induction of AD using minipumps to deliver NaN₃.

However, due to high cost, one-time usability and long treatment time, minipumps are unlikely to be useful tools for screening. A new technique was developed to provide an AD-like dementia model by selective cytochrome C complex inhibition using an intraperitoneal injection of NaN3 in different doses (10-15 mg/kg/day) in rats. The intraperitoneal injection dose-effect-interdependence study employed mitochondrial poison and developed a complex test system to research changes in cognitive functions induced by treatment with NaN3. It has been shown that NaN3 administration impairs learning and memory, and increases AChE levels in the brain ¹⁶⁹. NaN₃ was reported to cause oxidative damage that resulted in neuronal cell death. The progressive loss of neurons and necrosis were observed in the cortical and hippocampal areas of treated rats, thereby further indicating the potency of NaN3. These researchers confirmed that intraperitoneal injection of NaN₃ for 15 days resulted in a comparable level of dementia using implanted osmotic minipumps in rats, ultimately suggesting the involvement of NaN3 in inducing neurodegenerative disorder 185.

A growing body of knowledge has reported that oxidative stress (Figure 1) is a trigger, as well as one of the critical molecular changes for AD commencement¹⁸⁶. Oxidative stress may be caused by inhibition of mitochondrial function. The mitochondrial enzyme, Cytochrome oxidase (COX), has long been established to be predominantly implicated in aerobic energy metabolism and mitochondrial tasks in AD patients¹⁸⁷. Brains of AD patients have shown some mitochondrial abnormalities, particularly of CoX type. The primary toxic effect of NaN₃ has been identified as decreasing the action of CoX in the mitochondrial electron transport chain¹⁸⁵.

LPS-induced AD-like symptoms

LPS has been known to be used in various experiments, both in vitro and in vivo models of amyloidosis and neuroinflammation^{188,189}. A variety of neurodegenerative diseases, namely AD¹⁸⁹, Parkinson's disease¹⁹⁰, amyotrophic lateral sclerosis¹⁹¹, and multiple sclerosis 192, have been modeled using LPSinduced systemic inflammation (Figure 2). LPS is extracted from the external membrane of gram-negative bacteria. It has been reported as a strong endotoxin with resistivity to degradation by mammalian enzymes, therefore resulting in continual inflammatory stimuli¹⁹³, which produce proinflammatory cytokines. These proinflammatory cytokines stimulate both neuroimmune and neuroendocrine systems¹⁹⁴ that lead to virtually similar feedback formed by behavioral stress¹⁹⁵. Inflammation has been reported to be crucial in AD pathogenesis. However, the exact mechanisms through which it participates remains to be elucidated. Inflammatory proteins in the blood, two of which are C reactive protein (CRP) and IL6, have been reported to increase several years before the onset of clinical dementia in different studies^{12,196}. Inflammation is one of the factors that cause AD, although the link between infections and AD etiology has been an issue of debate for more than three decades¹⁹⁷. Previous studies have indicated that the expression levels of TNF- α , IL-1 β , and IL-6 within the hippocampus are upregulated in comparison with those of control following three days of LPS administration¹⁹⁸. Proinflammatory cytokines have already been found as the critical molecules which modulate immune responses; inability to reverse them during chronic inflammation would increase dyshomeostasis¹⁹⁸. Moreover, long-standing microglia activation which facilitates inflammatory mediators release, resulting in enhanced oxidative stress and nitrosative stress, is characterized by chronic inflammation¹⁹⁹.

Okadaic acid-induced memory impairment

Okadaic acid (OKA), a major polyether toxin, is a marine microalgae product that causes diarrhetic shellfish toxicity²⁰⁰. IC injection of OKA has been reported to induce memory decline in rats, therefore making it a valuable model agent to study for antidementia drug screening²⁰¹. In terms of mechanism of action, OKA has been found to be a restrictive and modest antagonist of serine/threonine phosphatases 1 and 2A^{202,203}, is associated with short- and longterm memory disruption in rats²⁰⁴, and triggers tau hyperphosphorylation (Figure 2) and neuronal cell death both in vivo²⁰⁵ and in vitro²⁰⁶. Previous findings have identified that OKA decreases basal synaptic transmission and inhibits the commencement of synaptic plasticity²⁰⁷. Also, OKA has been found to augment Ca²⁺ in a cultured hippocampal neuronal cells by ionotropic excitatory amino acid receptors, thereby leading to loss of neurons²⁰⁸.

Furthermore, research has demonstrated that OKA triggers the production of ROS in the hippocampus and reduces mitochondrial activity and mitochondrial potency, ultimately resulting in mitochondrial abnormalities in the brain of rats²⁰¹. Additionally, OKA has been known to inhibit phosphatases and cause hyperphosphorylation of proteins which subsequently leads to neuronal stress and ultimately to neurodegeneration²⁰¹. Previous findings revealed that OKA-mediated memory decline in rats is linked with exacerbated proinflammatory cytokines, such as TNF- α and IL-1, and with iNOS expression and total nitrite in the hippocampus and cortex²⁰⁹. Furthermore, infusion (bilateral) of OKA in the hippocampus produced spatial cognitive deficit as a result of raised GFAP expression, decreased GSH, and enhanced protein carbonylation and mitogen-activated protein kinase 38 (p38MAPK)²¹⁰. Several kinases, including MAPK, GSK-3 β and cyclin-dependent kinase 5 (Cdk5), have been identified to be implicated in the phosphorylation of tau protein at different positions seen in AD hyper-phosphorylated tau^{211,212}. Meanwhile, tau dephosphorylation is catalyzed through phosphatase PP2A and other phosphatases (PP1, 2B, and 5)^{213,214}. The precise blocking of PP2A by OKA could result in Alzheimer-like hyperphosphorylation and aggregation of tau in vivo²¹⁵ and in vitro^{216,217}. Memory loss due to OKA (intra-hippocampal) injection has been documented to be linked with apparent neuropathological changes, such as the formation of A β peptide-like structures, helical filamentlike phosphorylated tau, and neurodegeneration in the hippocampus²¹⁰. OKA is an essential means for



Figure 2: **Alzheimerogenic chemicals and some mechanisms used to induce AD.** The figure showed some insults underlying AD, the chemicals here induced AD-like cognitive impairments due to the individual mechanism of action. Chronic or acute exposure to any of these chemicals by the brain is associated with one or more of the pathophysiology contained in the box (dotted line). Adapted from *Lee et al.*, 2018¹⁸⁸.

analyzing²¹⁸ the cellular mechanisms involved by reversible protein phosphorylation during cell division, signal transduction, and formation of memory²¹⁷. At present, drugs that act by blocking tau phosphorylation are not available. Thus, this suggests that OKA could be a useful replacement tool towards unravelling therapeutic approaches for AD pathology¹⁹⁸.

CONCLUSION

Several chemicals have been established in order to unravel AD etiology and screen many possible therapeutic agents. This applies especially to aging, $A\beta$, aluminum, and D-galactose models, among others. Nonetheless, it is worthy to note that none of the available models recapitulates the exact pathology of AD, although every model is known to have its benefits and triggers a few of the pathophysiology linked with AD. Heavy metals, scopolamine, alcohol, and LPS are among the chemicals with proven neurotoxicity and have been used to induce AD-like cognitive impairment in rodents. Each chemical may act via specific mechanisms that differ in order to exacerbate AD pathogenesis. Thus, the dosage, as well as the time in which the chemical is administered in the AD models of rodents, may be higher or sometimes lesser to induce severe disease in human subjects. Importantly, other factors associated with the Alzheimerogenic chemicals are the route of administration, nature, duration (especially), as well as age and gender of the animals tested. This understanding is critical in order to design research that involves chemicallyinduced cognitive impairment.

ABBREVIATIONS

AChE: acetyl cholinesterase ACh: acetylcholine ChAT: choline acetyltransferase MDA: Melondialdehyde **SOD**: Superoxide dismutase iNOS: inducible nitric oxide **GSH**: glutathione GPx : glutathione peroxidase CAT : catalase DA: dopamine BDNF: brain derived neurotrophic factors NO: nitric oxide **ROS**: reactive oxygen species AC: total antioxidant content OG: oral gavage ICV: intracerebroventricular IH: intrahippocampal SC: subcutaneous SCM: Scopolamine

STZ: Streptozotocin **OKA**: Okadaic acid CLC: Colchicine SD: Sprague-Dawley **IP**: intraperitoneal ↑: increase \downarrow : decrease \leftrightarrow : unchaged LPS: Lippolyssacharide **TNF-** α : Tumor necrosis factor NFTs: neurofibrillary tangles COX-2: Cytocrome oxidase TBARS: Thiobarbituric acid reactive substance MPO: Myeloperoxidase GST: Glutathione S-transferase GFAP: Glial fibrillary acid protein

CONFLICTS OF INTEREST

Authors disclose none exists.

AUTHORS' CONTRIBUTIONS

All authors contributed to the design of the research. OM and SMC searched and summarized the data. SJ, MTBH, NHMN and MAMM reviewed and edited the first draft. All authors reviewed, commented and approved the final draft.

ACKNOWLEDGMENTS

The authors would like to acknowledge University Putra Malaysia for sponsoring this project (grant no.GP-IPS 9535400).

REFERENCES

- 1. of Economic United Nations D, Affairs PDS. World Population Ageing; 2015.
- Ravi SK, Ramesh BN, Mundugaru R, Vincent B. Multiple pharmacological activities of Caesalpinia crista against aluminium-induced neurodegeneration in rats: relevance for Alzheimer's disease. Environ Toxicol Pharmacol. 2018;58:202–11. PMID: 29408763. Available from: 10.1016/j. etap.2018.01.008.
- Tarragon E, Lopez D, Estrada C, Ana GC, Schenker E, Pifferi F, et al. Octodon degus: a model for the cognitive impairment associated with Alzheimer's disease. CNS Neurosci Ther. 2013;19(9):643–8. PMID: 23710760. Available from: 10.1111/cns.12125.
- Lam K, Pan K, Linnekamp JF, Medema JP, Kandimalla R. DNA methylation based biomarkers in colorectal cancer: A systematic review. Biochim Biophys Acta. 2016;1866(1):106–20. PMID: 27385266.
- Wani WY, Kandimalla RJ, Sharma DR, Kaushal A, Ruban A, Sunkaria A. Cell cycle activation in p21 dependent pathway: an alternative mechanism of organophosphateinduced dopaminergic neurodegeneration. Biochim Biophys Acta. 2016;1863(7):1858–1866. PMID: 27262357. Available from: 10.1016/j.bbadis.2016.05.014.
- Kandimalla R, Reddy PH. Multiple faces of dynamin-related protein 1 and its role in Alzheimer's disease pathogenesis. Biochim Biophys Acta. 2016;1862(4):814–28. PMID: 26708942. Available from: 10.1016/j.bbadis.2015.12.018.

- Nakagawa Y, Nakamura S, Kaśe Y, Noguchi T, Ishihara T. Colchicine lesions in the rat hippocampus mimic the alterations of several markers in Alzheimer's disease. Brain Res. 1987;408(1-2):57–64. PMID: 2885069. Available from: 10. 1016/0006-8993(87)90358-1.
- Li X, Bao X, Wang R. Experimental models of Alzheimer's disease for deciphering the pathogenesis and therapeutic screening (Review). Int J Mol Med. 2016;37(2):271–83. PMID: 26676932. Available from: 10.3892/ijmm.2015.2428.
- Alzheimer's Association Report (2018) Alzheimer's disease facts and figures Alzheimer's & Dementia, 14:367-429; 2018.
- 10. Villemagne VL, Burnham S, Bourgeat P, Brown B, Ellis KA, Salvado O, et al. Amyloid β deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: a prospective cohort study. Lancet Neurol. 2013;12(4):357–67. PMID: 23477989. Available from: 10.1016/S1474-4422(13)70044-9.
- Katzman R. Editorial: the prevalence and malignancy of Alzheimer disease. A major killer. Arch Neurol. 1976;33(4):217–8. PMID: 1259639. Available from: 10.1001/ archneur.1976.00500040001001.
- Zakaria R, Yaacob WMW, Othman Z, Long I, Ahmad AH, Al-Rahbi B. Lipopolysaccharide-induced memory impairment in rats: a model of Alzheimer's disease. Physiol Res. 2017;66(4):553–65. PMID: 28406691.
- Love S. Oxidative stress in brain ischemia. Brain Pathol. 1999;9(1):119–31. PMID: 9989455. Available from: 10.1111/j. 1750-3639.1999.tb00214.x.
- Chin-Chan M, Navarro-Yepes J, Quintanilla-Vega B. Environmental pollutants as risk factors for neurodegenerative disorders: alzheimer and Parkinson diseases. Front Cell Neurosci. 2015;9:124. PMID: 25914621. Available from: 10.3389/ fncel.2015.00124.
- Ramachandran S, Sanjay S, Dhanaraju M. Antiamnesic effect of Piracetam potentiated with Emblica officinalis and Curcuma longa in aluminium induced neurotoxicity of Alzheimer's disease. Int J Adv Res (Indore). 2013;1:185–96.
- Kent BA, Mistlberger RE. Sleep and hippocampal neurogenesis: implications for Alzheimer's disease. Front Neuroendocrinol. 2017;45:35–52. PMID: 28249715. Available from: 10.1016/j.yfrne.2017.02.004.
- 17. Alistair B, Steve I. CLINICAL REVIEW: alzheimer's disease. BMJ. 2009;21(338):467–71.
- Alzheimer's Disease International World Alzheimer Report (2015) The Global Impact of Dementia: An analysis of prevalence, incidence, cost & trends. Pp 1-82; 2015.
- Prince M, Wimo A, Guerchet M, Ali GC, Wu YT, Prina M, et al. World Alzheimer report, the global impact of dementia. An analysis of prevalence, incidence, cost and trends. London: Alzheimer's Disease International; 2015.
- Luy M, Minagawa Y. Gender gaps life expectancy and proportion of life in poor health. Health Rep. 2014;25(12):12–9. PMID: 25517936.
- Wimo A, Guerchet M, Ali GC, Wu YT, Prina AM, Winblad B, et al. The worldwide costs of dementia 2015 and comparisons with 2010. Alzheimers Dement. 2017;13(1):1–7. PMID: 27583652. Available from: 10.1016/j.jalz.2016.07.150.
- Adlimoghaddam A, Roy B, Albensi BC. Future Trends and the Economic Burden of Dementia in Manitoba: Comparison with the Rest of Canada and the World. Neuroepidemiology. 2018;51(1-2):71–81. PMID: 29969786. Available from: 10.1159/000490414.
- Mohandas E, Rajmohan V, Raghunath B. Neurobiology of Alzheimer's disease. Indian J Psychiatry. 2009;51(1):55–61. PMID: 19742193. Available from: 10.4103/0019-5545.44908.
- Swerdlow RH, Khan SM. A mitochondrial cascade hypothesis blright for sporadic Alzheimer's disease. Med Hypotheses. 2004;63(1):8–20. PMID: 15193340. Available from: 10.1016/j. mehy.2003.12.045.
- Swerdlow RH, Kish SJ. Mitochondria in Alzheimer's disease. Int Rev Neurobiol. 2002;53:341–85. PMID: 12512346. Available from: 10.1016/S0074-7742(02)53013-0.

- Gibson GE, Sheu KFR, Blass JP. Abnormalities of mitochondrial enzymes in Alzheimer disease. Journal of Neural Transmission. 1998;105(8-9):855–870. Available from: 10.1007/ s007020050099.
- Parker WD, Parks JK. Cytochrome c oxidase in Alzheimer's disease brain: purification and characterization. Neurology. 1995;45(3 Pt 1):482–6. PMID: 7898701. Available from: 10. 1212/WNL.45.3.482.
- Ding Q, Martin S, Dimayuga E, Bruce-Keller AJ, Keller JN. LMP2 knock-out mice have reduced proteasome activities and increased levels of oxidatively damaged proteins. Antioxid Redox Signal. 2006;8(1-2):130–135. PMID: 16487046. Available from: 10.1089/ars.2006.8.130.
- Swerdlow RH. Mitochondria in cybrids containing mtDNA from persons with mitochondriopathies. J Neurosci Res. 2007;85(15):3416–28. PMID: 17243174. Available from: 10.1002/jnr.21167.
- Duan AR, Jonasson EM, Alberico EO, Li C, Scripture JP, Miller RA, et al. Interactions between tau and different conformations of tubulin: implications for tau function and mechanism. J Mol Biol. 2017;429(9):1424–38. PMID: 28322917. Available from: 10.1016/j.jmb.2017.03.018.
- Medina M, Avila J. The role of extracellular Tau in the spreading of neurofibrillary pathology. Front Cell Neurosci. 2014;8:113. PMID: 24795568. Available from: 10.3389/fncel. 2014.00113.
- Neselius S, Zetterberg H, Blennow K, Randall J, Wilson D, Marcusson J, et al. Olympic boxing is associated with elevated levels of the neuronal protein tau in plasma. Brain Inj. 2013;27(4):425–33. PMID: 23473386. Available from: 10.3109/02699052.2012.750752.
- Chirita CN, Congdon EE, Yin H, Kuret J. Triggers of fulllength tau aggregation: a role for partially folded intermediates. Biochemistry. 2005;44(15):5862–5872. Available from: doi.org/10.1021/bi0500123.
- Sahara N, Maeda S, Murayama M, Suzuki T, Dohmae N, Yen SH, et al. Assembly of two distinct dimers and higherorder oligomers from full-length tau. Eur J Neurosci. 2007;25(10):3020–3029. Available from: doi.org/10.1111/j. 1460-9568.2007.05555.x.
- Mondragón-Rodríguez S, Basurto-Islas G, Santa-Maria I, Mena R, Binder LI, Avila J. Cleavage and conformational changes of tau protein follow phosphorylation during Alzheimer's disease. Int J Exp Pathol. 2008;89(2):81–90. Available from: 10.1111/j.1365-2613.2007.00568.x.
- Lasagna-Reeves CA, Castillo-Carranza DL, Guerrero-Muñoz MJ, Jackson GR, Kayed R. Preparation and characterization of neurotoxic tau oligomers. Biochemistry. 2010;49(47):10039– 10041. Available from: 10.1021/bi1016233.
- Patterson KR, Remmers C, Fu Y, Brooker S, Kanaan NM, Vana L, et al. Characterization of prefibrillar tau oligomers in vitro and in Alzheimer disease. J Biol Chem. 2011;286(26):23063– 23076. Available from: 10.1074/jbc.M111.237974.
- Gerson JE, Castillo-Carranza DL, Kayed R. Advances in therapeutics for neurodegenerative tauopathies: moving toward the specific targeting of the most toxic tau species. ACS Chem Neurosci. 2014;5(9):752–69. PMID: 25075869. Available from: 10.1021/cn500143n.
- Guerrero-Muñoz MJ, Gerson J, Castillo-Carranza DL. Tau Oligomers: The Toxic Player at Synapses in Alzheimer's Disease. Front Cell Neurosci. 2015;9:464. PMID: 26696824. Available from: 10.3389/fncel.2015.00464.
- Clavaguera F, Bolmont T, Crowther RA, Abramowski D, Frank S, Probst A, et al. Transmission and spreading of tauopathy in transgenic mouse brain. Nat Cell Biol. 2009;11(7):909–13. PMID: 19503072. Available from: 10.1038/ncb1901.
- Holmes BB, Diamond MI. Prion-like properties of tau protein: the importance of extracellular tau as a therapeutic target. J Biol Chem. 2014;289(29):19855–19861. Available from: 10. 1074/jbc.R114.549295.
- 42. Giacobini E, Gold G. Alzheimer disease therapy moving from amyloid- β to tau. Nat Rev Neurol. 2013;9(12):677–86. PMID:

24217510. Available from: 10.1038/nrneurol.2013.223.

- R BM, B G, K F, J M, A S, J C, et al. Tau and Aβ imaging, CSF measures, and cognition in Alzheimer's disease. Sci Transl Med. 2016;11(8(338)):338ra66. Available from: 10. 1126/scitranslmed.aaf2362.
- Neve RL, Harris P, Kosik KS, Kurnit DM, Donlon TA. Identification of cDNA clones for the human microtubule-associated protein tau and chromosomal localization of the genes for tau and microtubule-associated protein 2. Brain Res. 1986;387(3):271–80. PMID: 3103857.
- Mujika JI, Rezabal E, Mercero JM, Ruipérez F, Costa D, Ugalde JM, et al. Aluminium in biological environments: a computational approach. Comput Struct Biotechnol J. 2014;9(15):e201403002. PMID: 24757505. Available from: 10.5936/csbj.201403002.
- Goedert M, Spillantini MG, Jakes R, Rutherford D, Crowther RA. Multiple isoforms of human microtubule-associated protein tau: sequences and localization in neurofibrillary tangles of Alzheimer's disease. Neuron. 1989;3(4):519–26. PMID: 2484340. Available from: 10.1016/0896-6273(89)90210-9.
- Himmler A, Drechsel D, Kirschner MW, Martin DW. Tau consists of a set of proteins with repeated C-terminal microtubule-binding domains and variable N-terminal domains. Mol Cell Biol. 1989;9(4):1381–8. PMID: 2498649. Available from: 10.1128/MCB.9.4.1381.
- Callahan LM, Vaules WA, Coleman PD. Progressive reduction of synaptophysin message in single neurons in Alzheimer disease. J Neuropathol Exp Neurol. 2002;61(5):384–95. PMID: 12025941. Available from: 10.1093/jnen/61.5.384.
- Patterson KR, Remmers C, Fu Y, Brooker S, Kanaan NM, Vana L, et al. Characterization of prefibrillar tau oligomers in vitro and in Alzheimer disease. J Biol Chem. 2011;286(26):23063– 23076. Available from: 10.1074/jbc.M111.237974.
- Fá M, Puzzo D, Piacentini R, Staniszewski A, Zhang H, Baltrons MA. Extracellular tau oligomers produce an immediate impairment of LTP and memory. Sci Rep. 2016;6(1):19393. Available from: 10.1038/srep19393.
- Hamdane M, Delobel P, Sambo AV, Smet C, Bégard S, Violleau A, et al. Neurofibrillary degeneration of the Alzheimer-type: an alternate pathway to neuronal apoptosis? Biochem Pharmacol. 2003;66(8):1619–25. PMID: 14555242. Available from: 10.1016/S0006-2952(03)00533-1.
- Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. Science. 2002;297(5580):353–6. PMID: 12130773. Available from: 10.1126/science.1072994.
- Skovronsky DM, Lee VM, Trojanowski JQ. Neurodegenerative diseases: new concepts of pathogenesis and their therapeutic implications. Annu Rev Pathol. 2006;1(1):151–70. PMID: 18039111. Available from: 10.1146/annurev.pathol.1.110304. 100113.
- 54. Golde ET, Eckmann BC, Younkin SG. ochemical detection of A β isoforms: implications for pathogenesis, diagnosis and treatment of Alzheimer's disease. Biochim Biophys Acta. 2000;26(1502(1)):172–87. Available from: 10.1016/S0925-4439(00)00043-0.
- Kirkitadze MD, Bitan G, Teplow DB. Paradigm shifts in Alzheimer's disease and other neurodegenerative disorders: the emerging role of oligomeric assemblies. J Neurosci Res. 2002;69(5):567–77. PMID: 12210822. Available from: 10.1002/inr.10328.
- Kayed R, Head E, Thompson JL, McIntire TM, Milton SC, Cotman CW, et al. Common structure of soluble amyloid oligomers implies common mechanism of pathogenesis. Science. 2003;300(5618):486–9. PMID: 12702875. Available from: 10.1126/science.1079469.
- Sanchez-Varo R, Trujillo-Estrada L, Sanchez-Mejias E, Torres M, Baglietto-Vargas D, Moreno-Gonzalez I, et al. Abnormal accumulation of autophagic vesicles correlates with axonal and synaptic pathology in young Alzheimer's mice hippocampus. Acta Neuropathol. 2012;123(1):53–70. PMID: 22020633. Available from: 10.1007/s00401-011-0896-x.

- Suh YH, Checler F. Amyloid precursor protein, presenilins, and α-synuclein: molecular pathogenesis and pharmacological applications in Alzheimer's disease. Pharmacol Rev. 2002;54(3):469–525. PMID: 12223532. Available from: 10. 1124/pr.54.3.469.
- Kang J, Lemaire HG, Unterbeck A, Salbaum JM, Masters CL, Grzeschik KH, et al. The precursor of Alzheimer's disease amyloid A4 protein resembles a cell-surface receptor. Nature. 1987;325(6106):733–6. PMID: 2881207. Available from: 10.1038/325733a0.
- 60. Small DH, McLean CA. Alzheimer's disease and the amyloid β protein: what is the role of amyloid? J Neurochem. 1999;73(2):443–9. PMID: 10428038. Available from: 10.1046/ j.1471-4159.1999.0730443.x.
- Evin G, Weidemann A. Biogenesis and metabolism of Alzheimer's disease Abeta amyloid peptides. Peptides. 2002;23(7):1285–97. PMID: 12128085. Available from: 10. 1016/S0196-9781(02)00063-3.
- Pasternak JJ. An introduction to human molecular genetics: mechanisms of inherited diseases. Hoboken (New Jersey): John Wiley and Sons; 2005. Available from: 10.1002/ 0471719188.
- Devi L, Ohno M. Mitochondrial dysfunction and accumulation of the β-secretase-cleaved C-terminal fragment of APP in Alzheimer's disease transgenic mice. Neurobiol Dis. 2012;45(1):417–24. PMID: 21933711. Available from: 10. 1016/j.nbd.2011.09.001.
- Francis PT, Palmer AM, Snape M, Wilcock GK. The cholinergic hypothesis of Alzheimer's disease: a review of progress. J Neurol Neurosurg Psychiatry. 1999;66(2):137–47. PMID: 10071091. Available from: 10.1136/jnnp.66.2.137.
- Terry AV, Buccafusco JJ. The cholinergic hypothesis of age and Alzheimer's disease-related cognitive deficits: recent challenges and their implications for novel drug development. J Pharmacol Exp Ther. 2003;306(3):821–7. PMID: 12805474. Available from: 10.1124/jpet.102.041616.
- Hasselmo ME, Anderson BP, Bower JM. Cholinergic modulation of cortical associative memory function. J Neurophysiol. 1992;67(5):1230–46. PMID: 1597709. Available from: 10.1152/jn.1992.67.5.1230.
- Fine A, Hoyle C, Maclean CJ, Levatte TL, Baker HF, Ridley RM. Learning impairments following injection of a selective cholinergic immunotoxin, ME20.4 IgG-saporin, into the basal nucleus of Meynert in monkeys. Neuroscience. 1997;81(2):331–43. PMID: 9300425. Available from: 10.1016/ S0306-4522(97)00208-X.
- Sarter M, Bruno JP. Cognitive functions of cortical acetylcholine: toward a unifying hypothesis. Brain Res Brain Res Rev. 1997;23(1-2):28–46. PMID: 9063585. Available from: 10.1016/S0165-0173(96)00009-4.
- Miranda MI, Bermúdez-Rattoni F. Reversible inactivation of the nucleus basalis magnocellularis induces disruption of cortical acetylcholine release and acquisition, but not retrieval, of aversive memories. Proc Natl Acad Sci USA. 1999;96(11):6478–82. PMID: 10339613. Available from: 10.1073/pnas.96.11.6478.
- Haam J, Yakel JL. Cholinergic modulation of the hippocampal region and memory function. J Neurochem. 2017;142:111–21. PMID: 28791706. Available from: 10.1111/ jnc.14052.
- Brinkman SD, Gershon S. Measurement of cholinergic drug effects on memory in Alzheimer's disease. Neurobiol Aging. 1983;4(2):139–45. PMID: 6355883. Available from: 10.1016/ 0197-4580(83)90038-6.
- Summers WK, Viesselman JO, Marsh GM, Candelora K. Use of THA in treatment of Alzheimer-like dementia: pilot study in twelve patients. Biol Psychiatry. 1981;16(2):145–53. PMID: 7225483.
- Manoharan S, Guillemin GJ, Abiramasundari RS, Essa MM, Akbar M, Akbar MD. The role of reactive oxygen species in the pathogenesis of Alzheimer's disease, Parkinson's dis-

ease, and Huntington's disease: a mini review. Oxid Med Cell Longev. 2016;2016:8590578. PMID: 28116038. Available from: 10.1155/2016/8590578.

- Chen R, Lai UH, Zhu L, Singh A, Ahmed M, Forsyth NR. Reactive oxygen species formation in the brain at different oxygen levels: the role of hypoxia inducible factors. Front Cell Dev Biol. 2018;6:132. PMID: 30364203. Available from: 10.3389/fcell.2018.00132.
- Butterfield DA, Griffin S, Munch G, Pasinetti GM. Amyloid beta-peptide and amyloid pathology are central to the oxidative stress and inflammatory cascades under which Alzheimer's disease brain exists. J Alzheimers Dis. 2002;4(3):193–201. PMID: 12226538. Available from: 10. 3233/JAD-2002-4309.
- Gibson GL, Allsop D, Austen BM. Induction of cellular oxidative stress by the beta-amyloid peptide involved in Alzheimer's disease. Protein Pept Lett. 2004;11(3):257–70. PMID: 15182227. Available from: 10.2174/0929866043407101.
- 77. Abdul HM, Sultana R, Keller JN, Clair DKS, Markesbery WR, Butterfield DA. Mutations in amyloid precursor protein and presenilin-1 genes increase the basal oxidative stress in murine neuronal cells and lead to increased sensitivity to oxidative stress mediated by amyloid beta-peptide (1-42), HO and kainic acid: implications for Alzheimer's disease. J Neurochem. 2006;96(5):1322–35. PMID: 16478525. Available from: 10.1111/j.1471-4159.2005.03647.x.
- Butterfield DA, Reed T, Newman SF, Sultana R. Roles of amyloid beta-peptide-associated oxidative stress and brain protein modifications in the pathogenesis of Alzheimer's disease and mild cognitive impairment. Free Radic Biol Med. 2007;43(5):658–77. PMID: 17664130. Available from: 10. 1016/j.freeradbiomed.2007.05.037.
- Sultana R, Butterfield DA. Redox proteomics studies of in vivo amyloid beta-peptide animal models of Alzheimer's disease: insight into the role of oxidative stress. Proteomics Clin Appl. 2008;2(5):685–96. PMID: 21136866. Available from: 10.1002/ prca.200780024.
- Cheignon C, Tomas M, Bonnefont-Rousselot D, Faller P, Hureau C, Collin F. Oxidative stress and the amyloid beta peptide in Alzheimer's disease. Redox Biol. 2018;14:450– 64. PMID: 29080524. Available from: 10.1016/j.redox.2017. 10.014.
- Teixeira J, Silva T, Andrade PB, Borges F. Alzheimer's disease and antioxidant therapy: how long how far? Curr Med Chem. 2013;20(24):2939–52. PMID: 23409717. Available from: 10. 2174/1871523011320240001.
- Persson T, Popescu BO, Cedazo-Minguez A. Oxidative stress in Alzheimer's disease: why did antioxidant therapy fail? Oxidative medicine and cellular longevity. 2014;2014:427318. Available from: 10.1155/2014/427318.
- Salter MW, Stevens B. Microglia emerge as central players in brain disease. Nat Med. 2017;23(9):1018–27. PMID: 28886007. Available from: 10.1038/nm.4397.
- Zhang B, Gaiteri C, Bodea LG, Wang Z, McElwee J, Podtelezhnikov AA, et al. Integrated systems approach identifies genetic nodes and networks in late-onset Alzheimer's disease. Cell. 2013;153(3):707–20. PMID: 23622250. Available from: 10.1016/j.cell.2013.03.030.
- Guerreiro R, Wojtas A, Bras J, Carrasquillo M, Rogaeva E, Majounie E, et al. TREM2 variants in Alzheimer's disease. N Engl J Med. 2013;368(2):117–27. PMID: 23150934. Available from: 10.1056/NEJMoa1211851.
- Song W, Hooli B, Mullin K, Jin SC, Cella M, Ulland TK, et al. Alzheimer's disease-associated TREM2 variants exhibit either decreased or increased ligand-dependent activation. Alzheimers Dement. 2017;13(4):381–7. PMID: 27520774. Available from: 10.1016/j.jalz.2016.07.004.
- Bolós M, Perea JR, Avila J. Alzheimer's disease as an inflammatory disease. Biomol Concepts. 2017;8(1):37–43. PMID: 28231054. Available from: 10.1515/bmc-2016-0029.

- Paolicelli RC, Bolasco G, Pagani F, Maggi L, Scianni M, Panzanelli P, et al. Synaptic pruning by microglia is necessary for normal brain development. Science. 2011;333(6048):1456– 8. PMID: 21778362. Available from: 10.1126/science.1202529.
- Hong S, Dissing-Olesen L, Stevens B. New insights on the role of microglia in synaptic pruning in health and disease. Curr Opin Neurobiol. 2016;36:128–34. PMID: 26745839. Available from: 10.1016/j.conb.2015.12.004.
- Bliss TV, Collingridge GL, Morris RG. Synaptic plasticity in health and disease: introduction and overview. Philos Trans R Soc Lond B Biol Sci. 2013;369(1633):20130129. PMID: 24298133. Available from: 10.1098/rstb.2013.0129.
- Chen J, Sun Z, Jin M, Tu Y, Wang S, Yang X, et al. Inhibition of AGEs/RAGE/Rho/ROCK pathway suppresses non-specific neuroinflammation by regulating BV2 microglial M1/M2 polarization through the NF-kB pathway. J Neuroimmunol. 2017;305:108–14. PMID: 28284330. Available from: 10.1016/ j.jneuroim.2017.02.010.
- Hirbec HE, Noristani HN, Perrin FE. Microglia Responses in Acute and Chronic Neurological Diseases: What Microglia-Specific Transcriptomic Studies Taught (and did Not Teach) Us. Front Aging Neurosci. 2017;9:227. PMID: 28785215. Available from: 10.3389/fnagi.2017.00227.
- Baruch K, Deczkowska A, Rosenzweig N, Tsitsou-Kampeli A, Sharif AM, Matcovitch-Natan O, et al. PD-1 immune checkpoint blockade reduces pathology and improves memory in mouse models of Alzheimer's disease. Nat Med. 2016;22(2):135–7. PMID: 26779813. Available from: 10.1038/ nm.4022.
- Saresella M, Calabrese E, Marventano I, Piancone F, Gatti A, Calvo MG, et al. PD1 negative and PD1 positive CD4+ T regulatory cells in mild cognitive impairment and Alzheimer's disease. J Alzheimers Dis. 2010;21(3):927–38. PMID: 20634592. Available from: 10.3233/JAD-2010-091696.
- S M, C E, M I, P F, G A, F E. A potential role for the PD1/PD-L1 pathway in the neuroinflammation of Alzheimer's disease. Neurobiol Aging. 2012;33(3):e11–22.
- Jevtic S, Sengar AS, Salter MW, McLaurin J. The role of the immune system in Alzheimer disease: etiology and treatment. Ageing Res Rev. 2017;40:84–94. PMID: 28941639. Available from: 10.1016/j.arr.2017.08.005.
- McGeer PL, McGeer EG. Targeting microglia for the treatment of Alzheimer's disease. Expert Opin Ther Targets. 2015;19(4):497–506. PMID: 25435348. Available from: 10. 1517/14728222.2014.988707.
- Salter MW, Stevens B. Microglia emerge as central players in brain disease. Nat Med. 2017;23(9):1018–27. PMID: 28886007. Available from: 10.1038/nm.4397.
- More SV, Kumar H, Cho DY, Yun YS, Choi DK. Toxin-Induced Experimental Models of Learning and Memory Impairment. Int J Mol Sci. 2016;17(9):1447. PMID: 27598124. Available from: 10.3390/ijms17091447.
- Nallagouni CS, Reddy KP. Aluminium and fluoride impacts cortex, hippocampus and dentate gyrus structure in rats: protective effect of resveratrol. Int J Appl Biol Pharm Technol. 2017;8:89–97.
- Chiroma SM, Moklas MAM, Taib CNM, Baharuldin MT, Amon Z. d-galactose and aluminium chloride induced rat model with cognitive impairments. Biomed Pharmacother. 2018;103:1602–8. PMID: 29864948. Available from: 10.1016/ j.biopha.2018.04.152.
- Exley C. Human exposure to aluminium. Environ Sci Process Impacts. 2013;15(10):1807–16. PMID: 23982047. Available from: 10.1039/C3EM00374D.
- 103. Han S, Lemire J, Appanna VP, Auger C, Castonguay Z, Appanna VD. How aluminum, an intracellular ROS generator promotes hepatic and neurological diseases: the metabolic tale. Cell Biol Toxicol. 2013;29(2):75–84. PMID: 23463459. Available from: 10.1007/s10565-013-9239-0.
- Calvin C. Systematic review of potential health risks posed by pharmaceutical, occupational and consumer exposures to metallic and nanoscale aluminium, aluminium oxides, alu-

minium hydroxide and its soluble salts. Crit Rev Toxicol. 2014;44(sup4):1–80. Available from: 10.3109/10408444.2014. 934439.

- Bubser M, Byun N, Wood MR, Jones CK. Muscarinic receptor pharmacology and circuitry for the modulation of cognition. Handb Exp Pharmacol. 2012;208(208):121–66. PMID: 22222698. Available from: 10.1007/978-3-642-23274-9 7.
- Flicker C, Serby M, Ferris SH. Scopolamine effects on memory, language, visuospatial praxis and psychomotor speed. Psychopharmacology (Berl). 1990;100(2):243–50. PMID: 2305013. Available from: 10.1007/BF02244414.
- Spinks AB, Wasiak J, Villanueva EV, Bernath V. Scopolamine (hyoscine) for preventing and treating motion sickness. Cochrane Database Syst Rev. 2007;3(3):002851. PMID: 17636710. Available from: 10.1002/14651858.CD002851. pub3.
- Riedel G, Kang SH, Choi DY, Platt B. Scopolamine-induced deficits in social memory in mice: reversal by donepezil. Behav Brain Res. 2009;204(1):217–25. PMID: 19527754. Available from: 10.1016/j.bbr.2009.06.012.
- Pattipati KAS, Singh A. Animal Models in Drug Discovery of Alzheimer's Disease: A Mini Review. EC Pharmacology and Toxicology. 2016;2(1):60–79.
- 110. Klinkenberg I, Blokland A. The validity of scopolamine as a pharmacological model for cognitive impairment: a review of animal behavioral studies. Neurosci Biobehav Rev. 2010;34(8):1307–50. PMID: 20398692. Available from: 10. 1016/j.neubiorev.2010.04.001.
- 111. McEvoy GK. AHFS drug information. Scopolamine. Bethesda (MD): American Society of Health-System Pharmacists; 2005.
- 112. Khakpai F, Nasehi M, Haeri-Rohani A, Eidi A, Zarrindast MR. Scopolamine induced memory impairment; possible involvement of NMDA receptor mechanisms of dorsal hippocampus and/or septum. Behav Brain Res. 2012;231(1):1– 10. PMID: 22421366. Available from: 10.1016/j.bbr.2012.02. 049.
- 113. Singh P, Konar A, Kumar A, Srivas S, Thakur MK. Hippocampal chromatin-modifying enzymes are pivotal for scopolamineinduced synaptic plasticity gene expression changes and memory impairment. J Neurochem. 2015;134(4):642–51. PMID: 25982413. Available from: 10.1111/jnc.13171.
- 114. Portero-Tresserra M, Olmo ND, Martí-Nicolovius M, Guillazo-Blanch G, Vale-Martínez A. D-cycloserine prevents relational memory deficits and suppression of long-term potentiation induced by scopolamine in the hippocampus. Eur Neuropsychopharmacol. 2014;24(11):1798–807. PMID: 25453488. Available from: 10.1016/j.euroneuro.2014.10.002.
- 115. Blokland A, Honig W, Raaijmakers WG. Effects of intrahippocampal scopolamine injections in a repeated spatial acquisition task in the rat. Psychopharmacology (Berl). 1992;109(3):373–6. PMID: 1365638. Available from: 10.1007/ BF02245886.
- Elvander E, Schött PA, Sandin J, Bjelke B, Kehr J, Yoshitake T, et al. Intraseptal muscarinic ligands and galanin: influence on hippocampal acetylcholine and cognition. Neuroscience. 2004;126(3):541–57. PMID: 15183504. Available from: 10. 1016/j.neuroscience.2004.03.058.
- 117. Rogers JL, Kesner RP. Cholinergic modulation of the hippocampus during encoding and retrieval. Neurobiol Learn Mem. 2003;80(3):332–42. PMID: 14521875. Available from: 10.1016/S1074-7427(03)00063-7.
- 118. Wu YY, Wang X, Tan L, Liu D, Liu XH, Wang Q, et al. Lithium attenuates scopolamine-induced memory deficits with inhibition of GSK-3 β and preservation of postsynaptic components. J Alzheimers Dis. 2013;37(3):515–27. PMID: 23948897. Available from: 10.3233/JAD-130521.
- 119. Kumar A, Seghal N, Padi SV, Naidu PS. Differential effects of cyclooxygenase inhibitors on intracerebroventricular colchicine-induced dysfunction and oxidative stress in rats. Eur J Pharmacol. 2006;551(1-3):58–66. PMID: 17027965. Available from: 10.1016/j.ejphar.2006.08.076.

- Kumar A, Dogra S, Prakash A. Neuroprotective effects of Centella asiatica against intracerebroventricular colchicineinduced cognitive impairment and oxidative stress. Int J Alzheimers Dis. 2009;2009:972178. PMID: 20798885. Available from: 10.4061/2009/972178.
- 121. Evrard PA, Ragusi C, Boschi G, Verbeeck RK, Scherrmann JM. Simultaneous microdialysis in brain and blood of the mouse: extracellular and intracellular brain colchicine disposition. Brain Res. 1998;786(1-2):122–7. PMID: 9554978. Available from: 10.1016/S0006-8993(97)01454-6.
- 122. Ganguly R, Guha D. Alteration of brain monoamines & amp; EEG wave pattern in rat model of Alzheimer's disease & amp; protection by Moringa oleifera. Indian J Med Res. 2008;128(6):744–51. PMID: 19246799.
- 123. Saini N, Singh D, Sandhir R. Neuroprotective effects of Bacopa monnieri in experimental model of dementia. Neurochem Res. 2012;37(9):1928–37. PMID: 22700087. Available from: 10.1007/s11064-012-0811-4.
- 124. Subbaramaiah K, Hart JC, Norton L, Dannenberg AJ. Microtubule-interfering agents stimulate the transcription of cyclooxygenase-2. Evidence for involvement of ERK1/2 AND p38 mitogen-activated protein kinase pathways. J Biol Chem. 2000;275(20):14838–45. PMID: 10809726. Available from: 10.1074/jbc.275.20.14838.
- Kumar A, Dogra S, Prakash A. Protective effect of naringin, a citrus flavonoid, against colchicine-induced cognitive dysfunction and oxidative damage in rats. J Med Food. 2010;13(4):976–84. PMID: 20673063. Available from: 10. 1089/jmf.2009.1251.
- 126. Yu Z, Cheng G, Hu B. Mechanism of colchicine impairment on learning and memory, and protective effect of CGP36742 in mice. Brain Res. 1997;750(1-2):53–8. PMID: 9098529. Available from: 10.1016/S0006-8993(96)01158-4.
- 127. Sharma B, Singh N. Pitavastatin and 4'-hydroxy-3'methoxyacetophenone (HMAP) reduce cognitive dysfunction in vascular dementia during experimental diabetes. Curr Neurovasc Res. 2010;7(3):180–91. PMID: 20560881. Available from: 10.2174/156720210792231831.
- Raghavendra M, Maiti R, Kumar S, Acharya S. Role of aqueous extract of Azadirachta indica leaves in an experimental model of Alzheimer's disease in rats. Int J Appl Basic Med Res. 2013;3(1):37–47. PMID: 23776838. Available from: 10.4103/2229-516X.112239.
- 129. Tota S, Nath C, Najmi AK, Shukla R, Hanif K. Inhibition of central angiotensin converting enzyme ameliorates scopolamine induced memory impairment in mice: role of cholinergic neurotransmission, cerebral blood flow and brain energy metabolism. Behav Brain Res. 2012;232(1):66–76. PMID: 22460064. Available from: 10.1016/j.bbr.2012.03.015.
- Neha RK, Sodhi RK, Jaggi AS, Singh N. Animal models of dementia and cognitive dysfunction. Life Sci. 2014;109(2):73– 86. PMID: 25066372. Available from: 10.1016/j.lfs.2014.05. 017.
- 131. Zhou S, Yu G, Chi L, Zhu J, Zhang W, Zhang Y, et al. Neuroprotective effects of edaravone on cognitive deficit, oxidative stress and tau hyperphosphorylation induced by intracerebroventricular streptozotocin in rats. Neurotoxicology. 2013;38:136–45. PMID: 23932983. Available from: 10.1016/j.neuro.2013.07.007.
- 132. Yang S, Zhou G, Liu H, Zhang B, Li J, Cui R, et al. Protective effects of p38 MAPK inhibitor SB202190 against hippocampal apoptosis and spatial learning and memory deficits in a rat model of vascular dementia. BioMed Res Int. 2013;2013:215798. PMID: 24455679. Available from: 10.1155/2013/215798.
- Sodhi RK, Singh N, Jaggi AS. Neuroprotective mechanisms of peroxisome proliferator-activated receptor agonists in Alzheimer's disease. Naunyn Schmiedebergs Arch Pharmacol. 2011;384(2):115–24. PMID: 21607645. Available from: 10.1007/s00210-011-0654-6.
- 134. Correia SC, Santos RX, Santos MS, Casadesus G, Lamanna JC, Perry G, et al. Mitochondrial abnormalities in a

streptozotocin-induced rat model of sporadic Alzheimer's disease. Curr Alzheimer Res. 2013;10(4):406–19. PMID: 23061885. Available from: 10.2174/1567205011310040006.

- 135. Sharma M, Briyal S, Gupta YK. Effect of alpha lipoic acid, melatonin and trans resveratrol on intracerebroventricular streptozotocin induced spatial memory deficit in rats. Indian J Physiol Pharmacol. 2005;49(4):395–402. PMID: 16579392.
- Kaddour T, Kharoubi O, TaOA, Hellal N, Im'ene B, Aoues A. Aluminium-Induced Acute Neurotoxicity in Rats: treatment with aqueous extract of Arthrophytum (Hammada scoparia). J Acute Dis. 2016;5(6):470–82. Available from: 10.1016/j.joad. 2016.08.028.
- 137. Hussien HM, Abd-Elmegied A, Ghareeb DA, Hafez HS, Ahmed HE, El-Moneam NA. Neuroprotective effect of berberine against environmental heavy metals-induced neurotoxicity and Alzheimer's-like disease in rats. Food Chem Toxicol. 2018;111:432–44. PMID: 29170048. Available from: 10.1016/ j.fct.2017.11.025.
- Borai IH, Ezz MK, Rizk MZ, Aly HF, El-Sherbiny M, Matloub AA, et al. Therapeutic impact of grape leaves polyphenols on certain biochemical and neurological markers in AlCl3-induced Alzheimer's disease. Biomed Pharmacother. 2017;93:837–51. PMID: 28715867. Available from: 10.1016/j.biopha.2017.07. 038.
- Rostami F, Javan M, Moghimi A, Haddad-Mashadrizeh A, Fereidoni M. Streptozotocin-induced hippocampal astrogliosis and insulin signaling malfunction as experimental scales for subclinical sporadic Alzheimer model. Life Sci. 2017;188:172–85. PMID: 28867578. Available from: 10.1016/ j.lfs.2017.08.025.
- 140. Moreira-Silva D, Carrettiero DC, Oliveira AS, Rodrigues S, Santos-Lopes JD, Canas PM, et al. Anandamide Effects in a Streptozotocin-Induced Alzheimer's Disease-Like Sporadic Dementia in Rats. Front Neurosci. 2018;12(12):653. PMID: 30333717. Available from: 10.3389/fnins.2018.00653.
- 141. AM EH, NS S, HM A, RS ED. Protective effects of gingerol on streptozotocin-induced sporadic Alzheimer's disease: emphasis on inhibition of β -amyloid, COX-2, alpha-, beta-secretases and APH1a. Scientific reports. 2017;7(1):2902. Available from: 10.1038/s41598-017-02961-[186].
- 142. Samy DM, Ismail CA, Nassra RA, Zeitoun TM, Nomair AM. Downstream modulation of extrinsic apoptotic pathway in streptozotocin-induced Alzheimer's dementia in rats: Erythropoietin versus curcumin. European journal of pharmacology. 2016;770:52–60.
- 143. Bassani TB, Turnes JM, Moura EL, Bonato JM, Cóppola-Segovia V, Zanata SM, et al. Effects of curcumin on short-term spatial and recognition memory, adult neurogenesis and neuroinflammation in a streptozotocin-induced rat model of dementia of Alzheimer's type. Behav Brain Res. 2017;335:41–54. PMID: 28801114. Available from: 10.1016/j. bbr.2017.08.014.
- Palle S, Neerat P. Quercetin nanoparticles attenuates scopolamine induced spatial memory deficits and pathological damages in rats. Bull Fac Pharm Cairo Univ. 2017;55(1):101– 6. Available from: 10.1016/j.bfopcu.2016.10.004.
- Imam A, Ajao MS, Ajibola MI, Amin A, Abdulmajeed WI, Lawal AZ, et al. Black seed oil ameliorated scopolamine-induced memory dysfunction and cortico-hippocampal neural alterations in male Wistar rats. Bull Fac Pharm Cairo Univ. 2016;54(1):49–57. Available from: 10.1016/j.bfopcu.2015.12. 005.
- 146. Zaki HF, Abd-El-Fattah MA, Attia AS. Naringenin protects against scopolamine-induced dementia in rats. Faculty of Pharmacy. Cairo University. 2014;52:15–25.
- 147. Lee GY, Lee C, Park GH, Jang JH. Amelioration of Scopolamine-Induced Learning and Memory Impairment by α -Pinene in C57BL/6 Mice. Evidence-Based Complementary and Alternative Medicine. 2017;2017:4926815.
- Riedel G, Kang SH, Choi DY, Platt B. Scopolamine-induced deficits in social memory in mice: reversal by donepezil. Behav Brain Res. 2009;204(1):217–25. PMID: 19527754. Avail-

able from: 10.1016/j.bbr.2009.06.012.

- 149. Sil S, Ghosh T. Etoricoxib inhibits peripheral inflammation and alters immune responses in intracerebroventricular colchicine injected rats. J Neuroimmunol. 2018;317:15–23. PMID: 29501081. Available from: 10.1016/j.jneuroim.2018.01. 018.
- 150. Kumar A, Dogra S, Prakash A. Neuroprotective Effects of Centella asiatica against Intracerebroventricular Colchicine-Induced Cognitive Impairment and Oxidative Stress. International Journal of Alzheimer's Disease. 2009;2009:972178. Available from: 10.4061/2009/972178.
- 151. Pitchaimani V, Arumugam S, Thandavarayan RA, Thiyagarajan MK, Aiyalu R, Sreedhar R, et al. Nootropic activity of acetaminophen against colchicine induced cognitive impairment in rats. J Clin Biochem Nutr. 2012;50(3):241–4. PMID: 22573928. Available from: 10.3164/jcbn.11-73.
- 152. Mohamed AR, Soliman GY, Ismail CA, Mannaa HF. Neuroprotective role of vitamin D3 in colchicineinduced Alzheimer's disease in rats. Alexandria Journal of Medicine. 2015;51(2):127–36. Available from: 10.1016/j.ajme.2014.05.005.
- 153. Sil S, Ghosh T, Ghosh R. NMDA receptor is involved in neuroinflammation in intracerebroventricular colchicineinjected rats. J Immunotoxicol. 2016;13(4):474–89. PMID: 26788903. Available from: 10.3109/1547691X.2015.1130760.
- 154. Costa AP, Tramontina AC, Biasibetti R, Batassini C, Lopes MW, Wartchow KM, et al. Neuroglial alterations in rats submitted to the okadaic acid-induced model of dementia. Behav Brain Res. 2012;226(2):420–7. PMID: 21982813. Available from: 10. 1016/j.bbr.2011.09.035.
- 155. Broetto N, Hansen F, Brolese G, Batassini C, Lirio F, Galland F, et al. Intracerebroventricular administration of okadaic acid induces hippocampal glucose uptake dysfunction and tau phosphorylation. Brain Res Bull. 2016;124:136–43. PMID: 27108544. Available from: 10.1016/j.brainresbull.2016.04. 014.
- 156. Kamat PK, Rai S, Swarnkar S, Shukla R, Nath C. Mechanism of synapse redox stress in Okadaic acid (ICV) induced memory impairment: role of NMDA receptor. Neurochem Int. 2014;76:32–41. PMID: 24984170. Available from: 10.1016/ j.neuint.2014.06.012.
- 157. Zhang H, Wang P, Hou H, Wen H, Zhou H, Gao F, et al. Histone Modification Is Involved in Okadaic Acid (OA) Induced DNA Damage Response and G2-M Transition Arrest in Maize. PLoS One. 2016;11(5):e0155852. PMID: 27196101. Available from: 10.1371/journal.pone.0155852.
- 158. Cui SQ, Wang Q, Zheng Y, Xiao B, Sun HW, Gu XL, et al. Puerarin protects against damage to spatial learning and memory ability in mice with chronic alcohol poisoning. Braz J Med Biol Res. 2015;48(6):515–22. PMID: 25831201. Available from: 10.1590/1414-431x20144250.
- Marshall SA, Geil CR, Nixon K. Prior Binge Ethanol Exposure Potentiates the Microglial Response in a Model of Alcohol-Induced Neurodegeneration. Brain Sci. 2016;6(2):16. PMID: 27240410. Available from: 10.3390/brainsci6020016.
- 160. Cruz C, Meireles M, Silva SM. Chronic ethanol intake induces partial microglial activation that is not reversed by long-term ethanol withdrawal in the rat hippocampal formation. Neurotoxicology. 2017;60:107–15. PMID: 28408342. Available from: 10.1016/j.neuro.2017.04.005.
- 161. Lakshmi K, Karishma SK, Chand JG, Reddy MM, Babu AN, Kumar NB. Neuroprotective Activity of Terminalia chebula retz against Ethanol Induced Cognitive Impairment and Oxidative Stress in Rats: Promising for Regimentation the Risk of Alzheimer's Disease. Int J Res Ayurveda Pharm. 2018;9(4):90– 2
- 162. Uddin MS, Asaduzzaman M. Neuroprotective Activity of Asparagus racemosus Linn. Against Ethanol- Induced Cognitive Impairment and Oxidative Stress in Rats Brain: Auspicious for Controlling the Risk of Alzheimer's Disease. J Alzheimers Dis Parkinsonism. 2016;6(4):1–10.

- 163. Lee B, Shim I, Lee H. Gypenosides Attenuate Lipopolysaccharide-Induced Neuroinflammation and Memory Impairment in Rats. Evid Based Complement Alternat Med. 2018;2018:4183670. PMID: 30018656. Available from: 10.1155/2018/4183670.
- 164. Hou Y, Xie G, Miao F, Ding L, Mou Y, Wang L, et al. Pterostilbene attenuates lipopolysaccharide-induced learning and memory impairment possibly via inhibiting microglia activation and protecting neuronal injury in mice. Prog Neuropsychopharmacol Biol Psychiatry. 2014;54:92–102. PMID: 24709550. Available from: 10.1016/j.pnpbp.2014.03.015.
- 165. Kim YE, Hwang CJ, Lee HP, Kim CS, Son DJ, Ham YW. Inhibitory effect of punicalagin on lipopolysaccharideinduced neuroinflammation, oxidative stress and memory impairment via inhibition of nuclear factor-kappaB. Neuropharmacology. 2017;117:21e32. Available from: 10.1016/j. neuropharm.2017.01.025.
- 166. Wang Y, Wang M, Xu M, Li T, Fan K, Yan T, et al. Nootkatone, a neuroprotective agent from Alpiniae Oxyphyllae Fructus, improves cognitive impairment in lipopolysaccharide-induced mouse model of Alzheimer's disease. Int Immunopharmacol. 2018;62:77–85. PMID: 29990697. Available from: 10.1016/j. intimp.2018.06.042.
- Choi DY. Obovatol attenuates LPS-induced memory impairments in mice via inhibition of NF-jB signaling pathway. Neurochem Int. 2013;60:68–77.
- Szabados T, Dul C, Majtényi K, Hargitai J, Pénzes Z, Urbanics R. A chronic Alzheimer's model evoked by mitochondrial poison sodium azide for pharmacological investigations. Behav Brain Res. 2004;154(1):31–40. PMID: 15302108. Available from: 10.1016/j.bbr.2004.01.016.
- Wong-Riley M, Antuono P, Ho KC, Egan R, Hevner R, Liebl W. Cytochrome oxidase in Alzheimer's disease: biochemical, histochemical, and immunohistochemical analyses of the visual and other systems. Vision research. 1997;37(24):3593– 3608.
- Bennett MC, Mlady GW, Kwon YH, Rose GM. Chronic in vivo sodium azide infusion induces selective and stable inhibition of cytochrome c oxidase. J Neurochem. 1996;66(6):2606–11.
 PMID: 8632188. Available from: 10.1046/j.1471-4159.1996. 66062606.x.
- 171. Spinetta MJ, Woodlee MT, Feinberg LM, Stroud C, Schallert K, Cormack LK, et al. Alcohol-induced retrograde memory impairment in rats: prevention by caffeine. Psychopharmacology (Berl). 2008;201(3):361–71. PMID: 18758756. Available from: 10.1007/s00213-008-1294-5.
- 172. Patil S, Tawari S, Mundhada D, Nadeem S. Protective effect of berberine, an isoquinoline alkaloid ameliorates ethanol-induced oxidative stress and memory dysfunction in rats. Pharmacol Biochem Behav. 2015;136:13–20. PMID: 26159088. Available from: 10.1016/j.pbb.2015.07.001.
- Mailliard WS, Diamond I. Recent advances in the neurobiology of alcoholism: the role of adenosine. Pharmacol Ther. 2004;101(1):39–46. PMID: 14729391. Available from: 10.1016/j.pharmthera.2003.10.002.
- 174. Cui SQ, Wang Q, Zheng Y, Xiao B, Sun HW, Gu XL, et al. Puerarin protects against damage to spatial learning and memory ability in mice with chronic alcohol poisoning. Braz J Med Biol Res. 2015;48(6):515–22. PMID: 25831201. Available from: 10.1590/1414-431x20144250.
- Zola SM, Squire LR, Teng E, Stefanacci L, Buffalo EA, Clark RE. Impaired recognition memory in monkeys after damage limited to the hippocampal region. J Neurosci. 2000;20(1):451– 63. PMID: 10627621. Available from: 10.1523/JNEUROSCI. 20-01-00451.2000.
- Ji C, Li Q, Aisa H, Yang N, Dong L. Gossypium herbaceam extracts attenuate IBO-induced excitotoxicity in rat hippocampus. J Alzheimer Dis. 2009;16:331–339. Available from: 10.3233/JAD-2009-0979.
- Clark D, Tuor UI, Thompson R, Institoris A, Kulynych A, Zhang X, et al. Protection against recurrent stroke with resveratrol: endothelial protection. PLoS One. 2012;7(10):e47792. PMID:

23082218. Available from: 10.1371/journal.pone.0047792.

- 178. Karthick C, Periyasamy S, Jayachandran KS, Anusuyadevi M. Intrahippocampal Administration of Ibotenic Acid Induced Cholinergic Dysfunction via NR2A/NR2B Expression: Implications of Resveratrol against Alzheimer Disease Pathophysiology. Front Mol Neurosci. 2016;9(28):28. PMID: 27199654. Available from: 10.3389/fnmol.2016.00028.
- Rattan AK, Tejwani GA. The neurotoxic actions of ibotenic acid on cholinergic and opioid peptidergic systems in the central nervous system of the rat. Brain Res. 1992;571(2):298–305. PMID: 1611500. Available from: 10. 1016/0006-8993(92)90668-Y.
- 180. Kumar A, Prakash A, Pahwa D. Galantamine potentiates the protective effect of rofecoxib and caffeic acid against intrahippocampal Kainic acid-induced cognitive dysfunction in rat. Brain Res Bull. 2011;85(3-4):158–68. PMID: 21439356. Available from: 10.1016/i.brainresbull.2011.03.010.
- Antzoulatos EG, Byrne JH. Learning insights transmitted by glutamate. Trends Neurosci. 2004;27(9):555–60. PMID: 15331238. Available from: 10.1016/j.tins.2004.06.009.
- Mattson MP. Pathways towards and away from Alzheimer's disease. Nature. 2004;430(7000):631–9. PMID: 15295589. Available from: 10.1038/nature02621.
- Steven H. Anesthetic implications of a Near-Lethal Sodium Azide Exposure Anesth. Analog. 2007;104(1):229–30. Available from: 10.1213/01.ane.0000249841.29868.b2.
- Neha A, Bindra CS, Jain UK. Investigations on Molecular Mechanism Involved in Neuroprotective Effect of Vitamin D against Sodium Azide Induced Alzheimer's Disease in Rats. World Journal of Pharmaceutical Research. 2016;5(6):1154– 72.
- Gao C, Chang P, Yang L, Wang Y, Zhu S, Shan H, et al. Neuroprotective effects of hydrogen sulfide on sodium azideinduced oxidative stress in PC12 cells. Int J Mol Med. 2018;41(1):242–50. PMID: 29115393.
- Cadonic C, Sabbir MG, Albensi BC. Mechanisms of mitochondrial dysfunction in Alzheimer's disease. Mol Neurobiol. 2016;53(9):6078–90. PMID: 26537901. Available from: 10.1007/s12035-015-9515-5.
- 187. Anaeigoudari A, Soukhtanloo M, Reisi P, Beheshti F, Hosseini M. Inducible nitric oxide inhibitor aminoguanidine, ameliorates deleterious effects of lipopolysaccharide on memory and long term potentiation in rat. Life Sci. 2016;158:22–30. PMID: 27341994. Available from: 10.1016/j.lfs.2016.06.019.
- Lee HJ, Park MK, Seo YR. Pathogenic Mechanisms of Heavy Metal Induced-Alzheimer's Disease. Toxicol Environ Health. 2018;10(1):1–10. Available from: 10.1007/s13530-018-0340-
- Whitton PS. Inflammation as a causative factor in the aetiology of Parkinson's disease. Br J Pharmacol. 2007;150(8):963– 76. PMID: 17339843. Available from: 10.1038/sj.bjp.0707167.
- 190. Zhao W, Xie W, Le W, Beers DR, He Y, Henkel JS, et al. Activated microglia initiate motor neuron injury by a nitric oxide and glutamate-mediated mechanism. J Neuropathol Exp Neurol. 2004;63(9):964–77. PMID: 15453095. Available from: 10.1093/jnen/63.9.964.
- 191. Walter S, Doering A, Letiembre M, Liu Y, Hao W, Diem R, et al. The LPS receptor, CD14, in experimental autoimmune encephalomyelitis and multiple sclerosis. Cell Physiol Biochem. 2006;17(3-4):167–72. PMID: 16543733. Available from: 10. 1159/000092078.
- Ohanian SH, Schwab JH. Persistence of group a streptococcal cell walls related to chronic inflammation of rabbit dermal connective tissue. J Exp Med. 1967;125(6):1137–48. PMID: 5337778. Available from: 10.1084/jem.125.6.1137.
- 193. Maitra U, Deng H, Glaros T, Baker B, Capelluto DG, Li Z, et al. Molecular mechanisms responsible for the selective and low-grade induction of proinflammatory mediators in murine macrophages by lipopolysaccharide. J Immunol. 2012;189(2):1014–23. PMID: 22706082. Available from: 10.4049/jimmunol.1200857.

- 194. Oitzl MS, van Oers H, Schöbitz B, de Kloet ER. Interleukin-1 beta, but not interleukin-6, impairs spatial navigation learning. Brain Res. 1993;613(1):160–3. PMID: 8348300. Available from: 10.1016/0006-8993(93)90468-3.
- Sochocka M, Zwolińska K, Leszek J. The infectious etiology of Alzheimer's disease. Curr Neuropharmacol. 2017;15(7):996– 1009. PMID: 28294067. Available from: 10.2174/ 1570159X15666170313122937.
- Daulatzai MA. Fundamental role of pan-inflammation and oxidative-nitrosative pathways in neuropathogenesis of Alzheimer's disease. Am J Neurodegener Dis. 2016;5(1):1–28. PMID: 27073740.
- 197. Ghosh S, Lertwattanarak R, Jde JG, Galeana JJ, Li J, Zamarripa F. NMDA receptor is involved in neuroinflammation in intracerebroventricular colchicine-injected rats. Journal of Immunotoxicology. 2015;13(4):474–489. Available from: 10.3109/1547691X.2015.1130760.
- Kamat PK, Rai S, Nath C. Okadaic acid induced neurotoxicity: an emerging tool to study Alzheimer's disease pathology. Neurotoxicology. 2013;37:163–72. PMID: 23688530. Available from: 10.1016/j.neuro.2013.05.002.
- Fischer R, Maier O. Interrelation of oxidative stress and inflammation in neurodegenerative disease: role of TNF. Oxidative medicine and cellular longevity. 2015;2015.
- Kamat PK, Tota S, Saxena G, Shukla R, Nath C. Okadaic acid (ICV) induced memory impairment in rats: a suitable experimental model to test anti-dementia activity. Brain Res. 2010;1309:66–74. PMID: 19883632. Available from: 10.1016/j.brainres.2009.10.064.
- 201. Kamat PK, Tota S, Shukla R, Ali S, Najmi AK, Nath C. Mitochondrial dysfunction: a crucial event in okadaic acid (ICV) induced memory impairment and apoptotic cell death in rat brain. Pharmacol Biochem Behav. 2011;100(2):311–9. PMID: 21893081. Available from: 10.1016/j.pbb.2011.08.019.
- Cohen P, Holmes CF, Tsukitani Y. Okadaic acid: a new probe for the study of cellular regulation. Trends Biochem Sci. 1990;15(3):98–102. PMID: 2158158. Available from: 10.1016/ 0968-0004(90)90192-E.
- Ishihara H, Martin BL, Brautigan DL, Karaki H, Ozaki H, Kato Y, et al. Calyculin A and okadaic acid: inhibitors of protein phosphatase activity. Biochem Biophys Res Commun. 1989;159(3):871–7. PMID: 2539153. Available from: 10.1016/ 0006-291X(89)92189-X.
- 204. Maidana M, Carlis V, Galhardi FG, Yunes JS, Geracitano LA, Monserrat JM, et al. Effects of microcystins over short- and long-term memory and oxidative stress generation in hippocampus of rats. Chem Biol Interact. 2006;159(3):223–34. PMID: 16413006. Available from: 10.1016/j.cbi.2005.12.001.
- 205. He J, Yamada K, Zou LB, Nabeshima T. Spatial memory deficit and neurodegeneration induced by the direct injection of okadaic acid into the hippocampus in rats. J Neural Transm (Vienna). 2001;108(12):1435–43. PMID: 11810406. Available from: 10.1007/s007020100018.
- Cagnoli CM, Kharlamov E, Atabay C, Uz T, Manev H. Apoptosis induced in neuronal cultures by either the phosphatase inhibitor okadaic acid or the kinase inhibitor staurosporine is attenuated by isoquinolinesulfonamides H-7, H-8, and H-9. J Mol Neurosci. 1996;7(1):65–76. PMID: 8835783. Available

from: 10.1007/BF02736849.

- Koss DJ, Hindley KP, Riedel G, Platt B. Modulation of hippocampal calcium signalling and plasticity by serine/threonine protein phosphatases. J Neurochem. 2007;102(4):1009–23. PMID: 17442047. Available from: 10.1111/j.1471-4159.2007.04579.x.
- Fernández MT, Zitko V, Gascón S, Torreblanca A, Novelli A. Neurotoxic effect of okadaic acid, a seafood-related toxin, on cultured cerebellar neurons. Ann N Y Acad Sci. 1993;679:260–9. PMID: 8099773. Available from: 10.1111/ j.1749-6632.1993.tb18306.x.
- Kamat PK, Tota S, Rai S, Swarnkar S, Shukla R, Nath C. A study on neuroinflammatory marker in brain areas of okadaic acid (ICV) induced memory impaired rats. Life Sci. 2012;90(19-20):713–20. PMID: 22480513. Available from: 10.1016/j.lfs. 2012.03.012.
- 210. Costa AP, Tramontina AC, Biasibetti R, Batassini C, Lopes MW, Wartchow KM, et al. Neuroglial alterations in rats submitted to the okadaic acid-induced model of dementia. Behav Brain Res. 2012;226(2):420–7. PMID: 21982813. Available from: 10. 1016/j.bbr.2011.09.035.
- 211. Liu M, Choi S, Cuny GD, Ding K, Dobson BC, Glicksman MA, et al. Kinetic studies of Cdk5/p25 kinase: phosphorylation of tau and complex inhibition by two prototype inhibitors. Biochemistry. 2008;47(32):8367–77. PMID: 18636751. Available from: 10.1021/bi800732v.
- 212. Lucas JJ, Hernández F, Gómez-Ramos P, Morán MA, Hen R, Avila J. Decreased nuclear beta-catenin, tau hyperphosphorylation and neurodegeneration in GSK-3beta conditional transgenic mice. EMBO J. 2001;20(1-2):27–39. PMID: 11226152. Available from: 10.1093/emboj/20.1.27.
- Arendt T, Holzer M, Brückner MK, Janke C, Gärtner U. The use of okadaic acid in vivo and the induction of molecular changes typical for Alzheimer's disease. Neuroscience. 1998;85(4):1337–40. PMID: 9681968. Available from: 10. 1016/S0306-4522(97)00697-0.
- Liu F, Grundke-Iqbal I, Iqbal K, Gong CX. Contributions of protein phosphatases PP1, PP2A, PP2B and PP5 to the regulation of tau phosphorylation. Eur J Neurosci. 2005;22(8):1942–50.
 PMID: 16262633. Available from: 10.1111/j.1460-9568.2005. 04391.x.
- Zhang Z, Simpkins JW. Okadaic acid induces tau phosphorylation in SH-SY5Y cells in an estrogen-preventable manner. Brain Res. 2010;1345:176–81. PMID: 20457142. Available from: 10.1016/j.brainres.2010.04.074.
- de-la Rosa MA, Silva I, Nilsen J, Pérez MM, García-Segura LM, Avila J, et al. Estradiol prevents neural tau hyperphosphorylation characteristic of Alzheimer's disease. Ann N Y Acad Sci. 2005;1052(1):210–24. PMID: 16024764. Available from: 10.1196/annals.1347.016.
- Zhang Z, Simpkins JW. An okadaic acid-induced model of tauopathy and cognitive deficiency. Brain Res. 2010;1359:233–46. PMID: 20807517. Available from: 10.1016/j.brainres.2010.08.077.
- Fernández JJ, Candenas ML, Souto ML, Trujillo MM, Norte M. Okadaic acid, useful tool for studying cellular processes. Curr Med Chem. 2002;9(2):229–62. PMID: 11860357. Available from: 10.2174/0929867023371247.