

Increased Neuron Specific Enolase in a Hypoxic Ischemia Rat Model

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ABSTRACT

Introduction: Acute brain injury caused by cerebral ischemia, either due to stroke or ischemic hypoxic-ischemic encephalopathy (HIE), represents a major neurological cause of death and disability worldwide. Neuronal cell injury has been shown to correlate with a significant increase in neuron-specific enolase (NSE) levels in *in vitro* studies. This study aims to measure NSE levels in the blood serum of rats with HIE. **Methods:** This study used an experimental post-test design in which occlusion of the right common carotid artery (CCA) was performed on Wistar rats, which were then placed in a hypoxic chamber and reperfused after 60 minutes (treatment group). Neurological scores were collected over the first 24 hours. After 48 hours, experimental animals were sacrificed and their serum NSE levels were measured using ELISA. Statistical analysis using a T-test was performed for independent samples. **Results:** A total of 16 male rats were included in the study. Neurological scores indicated that all HIE group rats experienced hemiparesis to varying degrees. NSE levels in the treatment group were significantly higher than in the control group ($p < 0.05$). The mean NSE level in the treatment group was higher than that in the other group. **Conclusion:** Consistent with *in vitro* studies, we found that NSE was higher in the HIE rat model than in controls. These data support future studies to assess the fitness of serum NSE as a biomarker of brain damage.

Key words: hypoxic ischemic encephalopathy, mortality, neuron specific enolase

INTRODUCTION

Acute brain injury after cerebral ischemia, due to stroke or ischemic hypoxic-ischemic encephalopathy (HIE), remains a major cause of mortality and morbidity worldwide¹. Stroke occurs in 13 per 100,000 children over the age of one month every year, whereas in newborns, 25–40 children per 100,000 experience a stroke each year². Similarly, HIE occurs in 2–9 per 1,000 babies per year in both developed and developing countries³. Most affected children sustain lasting physical and/or neurological disabilities^{4,5}.

The prognosis after brain damage can be assessed using clinical, neuroimaging and electrophysiological methods. However, applying these methods together is often challenging. Biochemical markers and particularly markers that could be sampled from blood could offer a widely accessible alternative. Enolase is a glycolytic enzyme that converts 2-phosphoglycerate to phosphoenolpyruvate⁶. Previous research has demonstrated that neuron-specific enolase (NSE), which can be measured in cerebrospinal fluid, is a sensitive biomarker of neuronal injury, making it a useful tool for nerve and cerebral injury prognoses^{7,8}. Greater levels of NSE have been observed in adult gray matter, whereas decreased levels of NSE have been found in white matter. The interaction of NSE

with many other molecules in the central nervous system, including membrane-bound and cytoplasmic molecules, also increases the likelihood that NSE helps to activate glial cells during neuronal injury^{9,10}. We sought to measure the level of NSE in the serum of HIE rats, to determine whether the elevation of soluble NSE that has been reported in CSF could also be detected in peripheral blood, which would have immediate clinical relevance.

METHODS

Design

This was an experimental randomized posttest-only control group design.

Time and setting

Research activities were carried out at the Research and Development Centre for Stem Cells, Universitas Airlangga in July, and August 2021. The adaptation phase was carried out for one week, while the treatment and symptom observation phase lasted 48 hours.

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Animal subjects

Experimental animals used were all healthy, six-month-old, male Wistar rats, a strain of *Rattus norvegicus*. Rats were divided into a control group and a treatment group. Animals in the control group underwent no experimental interventions. In the treatment group, the right common carotid artery (CCA) was ligated. The rats were then placed for 45 minutes in a hypoxic atmosphere chamber, with an ambient gas composition of 8% O₂ and 92% N₂¹¹⁻¹³. Following hypoxia, reperfusion was performed by opening the CCA ligation after 60 minutes. After 48 hours, the rats were sacrificed. NSE levels were measured in blood serum by ELISA, using NSE reagent E-ELOR0058 (Elabscience Biotechnology Inc., China). Plates were read using a SPECTROstar[®] Nano plate reader (BMG Labtech, Germany).

Hypoxic-ischemic rat model

We established a cerebral ischemia model according to the Vannucci method. Wistar rats were anaesthetized with xylazine 10%, injected intramuscularly. An incision was made in the midventral cervical area in the middle of the neck, on the upper edge of the sternum. This allowed access to the sternocleidomastoid muscle and the sternohyoid muscle. The sternocleidomastoid muscle deep inside the sternohyoid muscle was pulled slowly until the sternohyoid muscle appeared. The common carotid artery (CCA) is normally wrapped in fibrous connective tissue, which contains vagus nerves. The CCA was carefully separated from the attached tissue using ophthalmic forceps. The unilateral CCA was then bound twice with two stitches with 4-0 silk, next to each other, and then the surgical wound was closed with a suture. Reperfusion was performed by opening of the CCA ligation for two hours.

Neurology Score

Clinical evidence of brain ischemia was assessed with a neurological examination, using a six-point Longa score scale as follows: 0: no neurological deficit; 1: left forefoot fails to fully extend, indicating a mild focal neurologic deficit; 2: turns left, meaning a moderate focal neurologic deficit; 3: falls to the left, indicating a severe focal neurologic deficit; 4: does not run spontaneously and exhibits a lower level of consciousness; 5: death due to brain ischemia. If the animal's score was 0 or 5, it was excluded from the study¹⁴.

Ethical clearance

All animal protocols were approved by the Committee of Animal Care and Use, Faculty of

Veterinary Medicine, Airlangga University No: 2.KE.019.02.2018.

Statistics

Normality of the control and treatment group data distributions was tested using the Shapiro-Wilk test, followed by a t-test for independent sample analysis. The Statistical Package for the Social Sciences (SPSS) 21.0 software program was used for this analysis.

RESULTS

Twenty-two healthy male Wistar rats that had not been used in previous studies were selected for the experiment. The characteristics of these rats are shown in **Table 1**.

We evaluated animals phenotypically for evidence of brain ischemia by performing a neurological examination on each animal, using the six-point Longa score scale (see methods). We found that all HIE group rats experienced hemiparesis at different levels, as shown in **Table 2**.

The normality of the NSE variable data was tested using the Shapiro-Wilks test. NSE data in each group showed a normal distribution. Levene's test was used to test whether our two groups had homogeneous variances. We found that the NSE levels were significantly higher in the treatment group compared with the control group (**Table 3**).

It is worth noting that the mortality of rats during the experiment reached 27% (3 rats) each in the control group and treatment group. The exact cause of death is unknown, but a severe neurological disorder is suspected. A total of 16 rats completed the study.

DISCUSSION

Enolase, or 2-phospho-D-glycerate hydrolase, is an essential metabolic enzyme and is found in the cytoplasm. Enolase is one of the enzymes of the glycolytic pathway that converts glucose into pyruvate. Brain enolase consists only of alpha and gamma subunits. Gamma enolase is categorized as neuron-specific enolase (NSE) because of the distinction of its neurons¹⁵. Previous studies have shown that NSE is detectable at high levels in CSF after brain injury. One outstanding question not addressed by those studies is the level of NSE in the blood serum in brain injury. This study set out to answer that question by measuring NSE level from blood serum of HIE rats model. We used CCA ligation to induce hypoxic ischemia and measured NSE levels after reperfusion. We found that NSE levels were higher in the serum of the study group compared to controls. NSE is considered a precise marker for neuronal injury but not glial injury^{8,16}. It is also

Table 1: Characteristics of subjects

Bodyweight (BW)	Group	
	Control	HIE
BW baseline (g)		
Mean	203.4	206.4
Standard Deviation	4.307	5.476
BW 48 hours post Ligation (g)		
Mean	199.0	204.3
Standard Deviation	4.629	5.599

Table 2: Neurology score of the subjects in HIE groups

	HIE Group							
	1	2	3	4	5	6	7	8
No Rats	1	2	3	4	5	6	7	8
Neurology Score after 24 hours	1	2	2	2	1	1	1	1

Table 3: Comparison of NSE level in both groups

Group	N	Mean	SD	95%CI	Mean Difference	P
Control	8	0.946	0.093	0.300-0.715	0.185	0.004*
HIE	8	1.132	0.118	0.300-0.709		

*Significant P < 0.05 with T-test for independent sample

a good marker for brain ischemic animal models, including local infarct and global ischemia^{16,17}. Cerebrospinal fluid (CSF) NSE levels are correlated with ischemia duration and the size of cerebral infarct. We interpreted our serum results after ischemia to mean that the level of neuron death was higher in our experimental animals than in controls.

One previous study compared the NSE levels in mouse CSF before and until seven days after ligation. The NSE level was measured using a radioimmunoassay kit. In those experiments, consolidation of the damaged area on the frontal lobe was evaluated to measure the infarct volume. There was a correlation between CSF NSE levels on the third day postligation and the infarct volume¹⁸.

NSE has been used as a more global marker of ischemia in humans, e.g., after cardiac arrest, status epilepticus, or drowning^{16,18}. A second study reported that focal ischemia treatment efficacy can be evaluated using NSE as a marker. They investigated the correlation between NSE and infarct volume in test animals by ligating the carotid artery. Ten minutes after ligation, animals were given intraperitoneal nicardipine 1.2 mg/kg and then again at 8, 16 and

24 hours after ligation. The mice that were given nicardipine had a 19% smaller infarct volume than the ligated group that was not treated. When NSE levels were measured 24 hours after ligation, they found that NSE was three times higher in ligated animals than in a control group that was not ligated. Interestingly, animals that were ligated and also treated with nicardipine had a lower level of NSE than those that were ligated and untreated, with the difference being up to 50% lower at 24 hours, 42% at 48 hours and 59% at 72 hours after ligation. These data showed that NSE levels may be correlated with different outcomes from therapeutic intervention for stroke^{19,20}.

Earlier studies have reported that NSE can be measured not only from CSF but also from blood¹⁰. In one such study, serum NSE levels were measured in mice with status epilepticus (SE) induced by lithium pilocarpine, and in healthy mice at the ages of 1, 2, 3 and 4 weeks. Brain damage was defined using a scale of 0 for no damage and up to 5 for more than 50% cellular loss. The 1- and 2-week-old mice had higher baseline serum NSE (s-NSE) than the other mice. After induction of SE, the 1-week-old mice did not show an increase in s-NSE, and there was no histological evidence of damage. In the older mice, NSE increased

between 18.9 ± 0.8 ng/ml in 2-week-old mice (vs 11.5 ± 0.5 for the control group) and 35.8 ± 2.1 ng/ml in 3-week-old mice (vs 12.1 ± 0.8 for the control group). After SE, the s-NSE of the mice also increased from 5.4 ± 0.4 for the control group to 30.4 ± 1.3 . They found that serum NSE after HIE injury correlated with histological changes and suggested that further studies to validate the use of NSE as a peripheral readout for brain injury may be warranted²¹.

In the current study, we were unable to replicate their study parameters and assess the presence of neuronal damage in the brain, since visualization of damage to the brain could be constrained by the limitations of equipment at our location.

CONCLUSIONS

Induced HIE in a rat model led to detectable increase in serum levels of NSE. Our study suggests that serum NSE may be a promising biomarker of brain damage. However, due to the limitations of our study (sample size, brain damage histological correlation with serum findings), further exploration will be needed, in varied neuropathological conditions and with larger sample sizes.

Abbreviations

BW: body weight; **CCA:** common carotid artery; **CSF:** cerebro spinal fluid; **HIE:** hypoxic-ischemic encephalopathy; **NSE:** neuron-specific enolase; **SE:** status epilepticus; **s-NSE:** serum neuron-specific enolase

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Author's contributions

Gunawan design conceptual framework, data collection, analysis and interpretation, discussion and summary. Noviandi collected data, revised the manuscript and supervised. Samosir revised the manuscript, figures and tables editing and literature review. All authors discussed the results and to the final version of the manuscript.

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Availability of data and materials

Data and materials used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

This study was conducted in accordance with the amended Declaration of Helsinki. The institutional review board approved the study, and all participants provided written informed consent.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

REFERENCES

- Meloni BP. Pathophysiology and neuroprotective strategies in hypoxic-ischemic brain injury and stroke. *Brain Sciences*. 2017;7(8):110. PMID: 28829350. Available from: [10.3390/brainsci7080110](https://doi.org/10.3390/brainsci7080110).
- Rivkin MJ, Bernard TJ, Dowling MM, Amlie-Lefond C. Guidelines for urgent management of stroke in children. *Pediatric Neurology*. 2016;56:8–17. PMID: 26969237. Available from: [10.1016/j.pediatrneurol.2016.01.016](https://doi.org/10.1016/j.pediatrneurol.2016.01.016).
- Eghbalian F. Frequency of Hypoxic-Ischemic Encephalopathy among Hospitalized Neonates in West Iran. *Iranian Journal of Pediatrics*. 2010;20(2):244–5. PMID: 23056714.
- van Handel M, Swaab H, de Vries LS, Jongmans MJ. Long-term cognitive and behavioral consequences of neonatal encephalopathy following perinatal asphyxia: a review. *European Journal of Pediatrics*. 2007;166(7):645–54. PMID: 17426984. Available from: [10.1007/s00431-007-0437-8](https://doi.org/10.1007/s00431-007-0437-8).
- Prastiya IG, Risky VP, Mira I, Retno AS, Darto S, Erny P. Risk factor of mortality in Indonesian children with cerebral palsy. *The Journal of Medical Investigation*. 2018;65(1.2):18–20. PMID: 29593188. Available from: [10.2152/jmi.65.18](https://doi.org/10.2152/jmi.65.18).
- Yardimoğlu M, Ilbay G, Dalcik C, Dalcik H, Sahin D, Ates N. Immunocytochemistry of neuron specific enolase (NSE) in the rat brain after single and repeated epileptic seizures. *The International Journal of Neuroscience*. 2008;118(7):981–93. PMID: 18569155. Available from: [10.1080/00207450701769232](https://doi.org/10.1080/00207450701769232).
- León-Lozano MZ, Arnaez J, Valls A, Arca G, Agut T, Alarcón A. Cerebrospinal fluid levels of neuron-specific enolase predict the severity of brain damage in newborns with neonatal hypoxic-ischemic encephalopathy treated with hypothermia. *PLoS One*. 2020;15(6):e0234082. PMID: 32479533. Available from: [10.1371/journal.pone.0234082](https://doi.org/10.1371/journal.pone.0234082).
- Morozova AY, Arutjunyan AV, Milyutina YP, Morozova PY, Kozina LS, Zhuravin IA. Influence of prenatal hypoxia on the content of neuron specific enolase in the structures of the brain and blood serum of rats in early ontogeny. *Neurochemical Journal*. 2020;14(3):290–4. Available from: [10.1134/S1819712420030083](https://doi.org/10.1134/S1819712420030083).
- Lafon-Cazal M, Bougault I, Steinberg R, Pin JP, Bockaert J. Measurement of gamma-enolase release, a new method for selective quantification of neurotoxicity independently from glial lysis. *Brain Research*. 1992;593(1):63–8. PMID: 1458320. Available from: [10.1016/0006-8993\(92\)91264-F](https://doi.org/10.1016/0006-8993(92)91264-F).

10. Haque A, Polcyn R, Matzelle D, Banik NL. New insights into the role of Neuron-Specific Enolase in neuro-Inflammation, neurodegeneration, and neuroprotection. *Brain Sciences*. 2018;8(2):33. PMID: 29463007. Available from: [10.3390/brainsci8020033](https://doi.org/10.3390/brainsci8020033).
11. Northington FJ. Brief update on animal models of hypoxic-ischemic encephalopathy and neonatal stroke. *ILAR Journal*. 2006;47(1):32–8. PMID: 16391429. Available from: [10.1093/ilar.47.1.32](https://doi.org/10.1093/ilar.47.1.32).
12. Wilson MD. Wilson. Animal models of cerebral palsy: hypoxic brain injury in the newborn. *Iranian Journal of Child Neurology*. 2015;9(2):9–16. PMID: 26221157.
13. Vannucci RC, Vannucci SJ. Perinatal hypoxic-ischemic brain damage: evolution of an animal model. *Developmental Neuroscience*. 2005;27(2-4):81–6. PMID: 16046840. Available from: [10.1159/000085978](https://doi.org/10.1159/000085978).
14. Bachour SP, Hevesi M, Bachour O, Sweis BM, Mahmoudi J, Brekke JA. Comparisons between Garcia, Modo, and Longa rodent stroke scales: optimizing resource allocation in rat models of focal middle cerebral artery occlusion. *Journal of the Neurological Sciences*. 2016;364(364):136–40. PMID: 27084232. Available from: [10.1016/j.jns.2016.03.029](https://doi.org/10.1016/j.jns.2016.03.029).
15. Yardimoğlu M, Ilbay G, Dalcik C, Dalcik H, Sahin D, Ates N. Immunocytochemistry of neuron specific enolase (NSE) in the rat brain after single and repeated epileptic seizures. *The International Journal of Neuroscience*. 2008;118(7):981–93. PMID: 18569155. Available from: [10.1080/00207450701769232](https://doi.org/10.1080/00207450701769232).
16. Zetterberg H, Smith DH, Blennow K. Biomarkers of mild traumatic brain injury in cerebrospinal fluid and blood. *Nature Reviews Neurology*. 2013;9(4):201–10. PMID: 23399646. Available from: [10.1038/nrneuro.2013.9](https://doi.org/10.1038/nrneuro.2013.9).
17. Palmio J, Peltola J, Vuorinen P, Laine S, Suhonen J, Keränen T. Normal CSF neuron-specific enolase and S-100 protein levels in patients with recent non-complicated tonic-clonic seizures. *Journal of the Neurological Sciences*. 2001;183(1):27–31. PMID: 11166790. Available from: [10.1016/S0022-510X\(00\)00478-0](https://doi.org/10.1016/S0022-510X(00)00478-0).
18. Hatfield RH, McKernan RM. CSF neuron-specific enolase as a quantitative marker of neuronal damage in a rat stroke model. *Brain Research*. 1992;577(2):249–52. PMID: 1606499. Available from: [10.1016/0006-8993\(92\)90280-M](https://doi.org/10.1016/0006-8993(92)90280-M).
19. Pratamastuti D, Gunawan PI, Saharso D. Serum neuron specific enolase is increased in pediatric acute encephalitis syndrome. *Korean Journal of Pediatrics*. 2017;60(9):302–6. PMID: 29042874. Available from: [10.3345/kjp.2017.60.9.302](https://doi.org/10.3345/kjp.2017.60.9.302).
20. Kittaka M, Giannotta SL, Zelman V, Correale JD, DeGiorgio CM, Weiss MH. Attenuation of brain injury and reduction of neuron-specific enolase by nicardipine in systemic circulation following focal ischemia and reperfusion in a rat model. *Journal of Neurosurgery*. 1997;87(5):731–7. PMID: 9347982. Available from: [10.3171/jns.1997.87.5.0731](https://doi.org/10.3171/jns.1997.87.5.0731).
21. Sankar R, Shin DH, Wasterlain CG. Serum neuron-specific enolase is a marker for neuronal damage following status epilepticus in the rat. *Epilepsy Research*. 1997;28(2):129–36. PMID: 9267777. Available from: [10.1016/S0920-1211\(97\)00040-5](https://doi.org/10.1016/S0920-1211(97)00040-5).

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